



DIVERSITY OF SOIL ALGAE IN THE FARMLANDS OF KHASI HILLS, MEGHALAYA

*Carefulness M. Dirborne and Papiya Ramanujam

Algal Ecology Laboratory, Centre for Advance Studies in Botany, Department of Botany, School of Life Sciences, North-eastern Hill University, Shillong, Meghalaya, India-793022

ARTICLE INFO

Article History:

Received 16th January, 2018
Received in revised form
24th February, 2018
Accepted 20th March, 2018
Published online 30th April, 2018

Key Words:

Diversity, farmlands,
Fertiliser, Cyanobacteria,
Soil algae

ABSTRACT

The present paper deals with the structure of soil algal community in four farmlands located in Khasi Hills, Meghalaya. The chosen farmlands were corn field (*Zea mays* L.), rice field (*Oryzasativa* Linn.), vegetable farm (*Solanumtuberosum* L., *Lactucasativa* L. and *Brassica oleracea* var. *capitata* f. *alba*) and citrus plantation (*Citrusreticulata* Blanco.). A total of 158 algal taxa have been identified, out of which Chlorophyta+Xanthophyceae members (64 taxa) form the major group, followed by Bacillariophyceae (62 taxa) and Cyanobacteria (32 taxa). Species diversity was recorded higher in rice field (1.55- 2.62) followed by vegetable farm (0.69-2.10), corn field (1.05-2.01) and citrus plantation (0.98-1.99). Use of fertiliser, insecticides and pesticides adversely affected the composition and abundance of the algal community in corn field, vegetable field and citrus plantation.

Copyright © 2018, Carefulness M. Dirborne and Papiya Ramanujam. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Carefulness M. Dirborne and Papiya Ramanujam, 2018. "Diversity of soil algae in the farmlands of khasi hills, meghalaya", *International Journal of Development Research*, 8, (04), 20000-20006.

INTRODUCTION

Algae are ubiquitous and vary widely in many habitats. Soil is one of the most important non-aqueous ecosystems for algae (Zenova *et al.*, 1995). Unlike most other microorganisms, algae contain chlorophylls and are present in most of the soils where moisture and sunlight is available (Hoffman, 1989). In farmlands, soil algae and Cyanobacteria play important roles because they have tremendous potential to serve as sources of nitrogen and carbon for other organisms and improve soil fertility by adding organic matter (Mishra and Pabbi, 2004). They also help to stabilize the soil surface by reducing soil erosion (Hu *et al.*, 2004). Soil algae mainly Cyanobacteria are known to associate with plant roots and enhance the activities of host plant. The structure and function of soil biota get affected significantly by different agricultural practices. Application of pesticides, chemical fertilisers and agronomic practices affect plants, animal life and soil community structure (Paoletti *et al.*, 1988). Despite numerous studies on soil algae (Metting, 1981; Lukesova, 1993; Neustupa, 2001; Zancanet *et al.* 2006; Lin *et al.* 2013; Ray and Thomas, 2013; Vijayan, 2015), it is still difficult to draw general conclusions

on the diversity of the soil algae and their influence on ecosystem functions. Paoletti and Pimentel (1992) reported that changes in land management significantly influenced the soil biota resulting in certain transient or permanent signs in the system. Literature regarding role of soil algae are available from different parts of the world but information about the diversity and function of soil algae in Khasi Hills, Meghalaya is very limited. In the present study, a survey of soil algal diversity in different types of farmlands in Khasi Hills have been undertaken to established the importance of the soil algae in this region.

MATERIALS AND METHODS

Study sites: Four different farmlands (corn field, rice field, vegetable farm and citrus plantation) were selected in Khasi Hills, Meghalaya (Table 1).

Collection of soil samples: Soil samples were collected on a monthly basis from January 2016 to December 2016. 10 random sites were selected from each farmlands and 500 g of soil was collected from the upper layer (0-2 cm) by removing the surface litter at each site. About 10 g of each soil sample was placed in a flask and diluted 100-fold with distilled water. Different culture media like Bold's Basal Medium (BBM) and

*Corresponding author: Carefulness M. Dirborne,

Algal Ecology Laboratory, Centre for Advance Studies in Botany, Department of Botany, School of Life Sciences, North-eastern Hill University, Shillong, Meghalaya, India-793022

BG-11 were used. One ml of 0.11 N solution of $\text{Na}_2\text{SiO}_3 \cdot 9\text{H}_2\text{O}$ was added to each litre of BBM to provide silicon for diatoms (Sukala and Davis, 1994). The number of algae appeared in culture media were determined by counting algal colonies and estimated per 1g dry weight (d.w.) of soil. Colonies of Chlorophyta (CH) and Xanthophyceae (X) (hereinafter CH+X) were counted together, Cyanobacteria (CY) and Bacillariophyceae (D) were counted separately (Zancan *et al.*, 2006). Algae were observed under trinocular microscope (Olympus BX41) and identified to the possible lower taxonomic level with the help of standard books and Monographs like Fritsch (1935), Desikachary (1959), Philipose (1967), Prescott (1982), Gandhi (1998), and John *et al.* (2002). Taxonomy was updated using the online database Algae Base (Guiry and Guiry, 2017) and ADIAC (1999). Species diversity was calculated using the Shannon-Wiener diversity index (1963) following the formula,

$$H' = -\sum_{i=1}^s p_i \ln p_i$$

Where $p_i = n_i/N$, n_i = number of individuals of the i^{th} species; N = total number of individuals of all the species.

using ANOVA with Microsoft Office Excel. Principal Component Analysis (PCA) was employed to compare the farmlands with soil characteristics in the first analysis and a second analysis was conducted to compare the algal community characteristics on the abundance of taxa in the farmlands. The data were also subjected to Pearson's correlation analysis. These analysis were carried out in PAST software version 1.93.

RESULTS

Physico-chemical characteristics of soil: Physico-chemical parameters except for phosphorus varied significantly in the four farmlands ($p < 0.05$). The farmlands are acidic with pH ranging from 5.55 to 6.60. The lowest pH was measured for rice field while the highest for citrus plantation. Moisture content, organic carbon and total nitrogen were highest in rice field with 53.68%, 1.90 % and 0.22 % respectively and lowest in citrus plantation with 20.67%, 1.23% and 0.11% respectively. Conductivity recorded did not show much variation in all the farmlands. A value of $0.03\text{mS}^{-1}/\text{cm}$ was recorded for rice field, followed by $0.02\text{mS}^{-1}/\text{cm}$ for corn and vegetable farm and a value of $0.01\text{mS}^{-1}/\text{cm}$ was observed in citrus plantation.

Table 1. Sampling plots with their locations

Sampling plots	Village	Elevation (m a.s.l)	Latitude Longitude	Fertiliser/pesticides/ Insecticides application
Corn field (<i>Zea mays</i> L.)	Mairang	1720	25°32.713'N 91°39.486' E	Urea and FYM (Farm Yard Manure)
Rice field (<i>Oryzasativa</i> Linn.)	Sohiong	1737	25°30.440'N 91°42.043'E	Urea
Vegetable farms potato (<i>Solanumtuberosum</i> L.), lettuce (<i>Lactucasativa</i> L.) white cabbage (<i>Brassica oleracea</i> var. <i>capitata</i> f. <i>alba</i>)	Mylliem	1685	25°29.529'N, 91°48.724' E	NPK
Citrus plantation (<i>Citrusreticulata</i> Blanco.)	Mawlai	1406	25°35.214'N, 91°58.306'E	Occasional application of Insecticides and pesticides

Table 2. Physicochemical properties of soil in different farmlands of Khasi Hills

Parameters	Corn field	Rice field	Vegetable farm	Citrus Plantation	F-value	P-value
pH	6.32±0.59	5.55±0.35	5.86±0.44	6.60±0.25	7.19	0.0018*
Moisture content (%)	32.61±5.03	53.68±5.40	34.22±5.76	20.67±2.80	40.79	1.3E-10*
Conductivity (mS^{-1}/cm)	0.02±0.005	0.03±0.005	0.02±0.005	0.01±0.005	11.96	0.0001*
Organic carbon (%)	1.27±0.36	2.34±0.59	1.90±0.15	1.23±0.36	10.07	0.0002*
Total nitrogen (%)	1.18±0.07	1.76±0.24	1.10±0.16	0.11±0.02	117.33	7.28E-13*
Available Phosphorus (%)	0.0032±0.001	0.0038±0.00	0.0049±0.0	0.0056±0.001	0.57	0.64
Exchangeable Potassium (%)	0.02±0.005	0.02±0.005	0.03±0.01	0.08±0.008	24.48	6.7E-07*

*significant (P at 0.05 and 0.01 levels).

Soil analysis

Soil samples were soaked in deionized water (1 water: 5 soil) to prepare a soil solution for the measurements of pH and electrical conductivity. pH and conductivity were read using digital pH meter and conductivity meter respectively. Moisture content of the soil was determined by oven dry method (drying 10 grams of soil in a hot air oven at 105°C). Soil organic carbon was estimated following the method by Walkley and Black, (1934) while total nitrogen, phosphorus and potassium were estimated as per Jackson (1967).

Statistical Analyses

Data for different physico-chemical parameters and algal abundance were obtained on the basis of mean of ten independent samples. The data were statistically analysed

Phosphorus was observed to be highest in citrus plantation with 0.0056% and lowest in corn with 0.0032%. In case of potassium, citrus plantation was recorded with a higher value of 0.08% as compared to lower value of 0.02% in corn and rice fields (Table 2).

Soil algal community structure: Based on morphology, a total of 158 taxa were identified from four different farmlands. The highest number of taxa belonged to CH+X group (64 taxa) followed by Bacillariophyceae (62 taxa) and then the Cyanobacteria (32 taxa). Out of 158 species, 71 taxa were recorded from rice field, 58 taxa in vegetable farms, 35 taxa in citrus plantation and 30 taxa from corn field (Table 3). Algal species diversity was richer in rice field and vegetable farms. Members of green algae were recorded in high number in vegetable farms while those of Bacillariophyceae and Cyanobacteria were higher in rice field. Diversity index was

Table 3. List of algal taxa recorded in four farmlands of Khasi Hills

Taxa	Corn field	Rice field	Vegetable farm	Citrus Plantation
Cyanobacteria				
<i>Anabaena catenula</i> Kützing ex Bornet&Flahault		+		
<i>Anabaena constricta</i> (Szafer) Geitler			+	
<i>Anabaena spiroides</i> Klebahn	+			+
<i>Anabaena variabilis</i> Kützing ex Bornet&Flahault	+	+	+	+
<i>Anabaenopsis</i> sp		+		
<i>Aphanocapsaelachista</i> West &G.S.West			+	
<i>Chroococcus minimus</i> (Keissler) Lemmermann				+
<i>Cylindrospermum michailovskoense</i> Elenkin		+		
<i>Cylindrospermum muscicola</i> Kützing ex Bornet&Flahault		+		
<i>Leptolyngbya</i> sp				
<i>Lyngbyadendrobya</i> Brühl&Biswas		+		
<i>Lyngbyapalmarum</i> Brühl&Biswas			+	
<i>Lyngbyasemiplena</i> J. Agardh ex Gomont			+	
<i>Lyngbyaspiralis</i> Geitler			+	
<i>Merismopediasp</i>				+
<i>Microcystis smithii</i> Komarek & Anagnostidis		+		
<i>Nostoc carneum</i> C. Agardh ex Bornet & Flahault		+	+	+
<i>Nostoc commune</i> Vaucher ex Bornet & Flahault			+	
<i>Nostoc muscorum</i> C. Agardh ex Bornet&Flahault	+			
<i>Nostoc pruneforme</i> C. Agardh ex Bornet&Flahault			+	
<i>Oscillatoria curviceps</i> C. Agardh ex Gomont	+			
<i>Oscillatoria germinata</i> Schwabe ex Gomont		+		
<i>Oscillatoria limosa</i> C. Agardh ex Gomont	+	+		
<i>Oscillatoria nigra</i> Vaucher ex Gomont		+		
<i>Oscillatoria rubescens</i> De Candolle ex Gomont		+		
<i>Oscillatoriasp</i>		+		+
<i>Oscillatoria tenuis</i> C. Agardh ex Gomont				+
<i>Phormidium abronema</i> Skuja		+		
<i>Phormidium undatum</i> Kützing ex Gomont			+	
<i>Phormidium tenue</i> Gomont			+	
<i>Stigonema ocellatum</i> Thuret ex Bornet & Flahault		+		
<i>Synechococcus aeruginosus</i> Nägeli		+	+	+
Bacillariophyceae				
<i>Achnanthes lanceolata</i> (Brébisson ex Kützing) Grunow				+
<i>Achnantheidium mexiguum</i> (Grunow) Czarnecki				+
<i>Amphora elliptica</i> (Agardh) Kützing			+	
<i>Craticula subhalophila</i> (Hustedt) Lange-Bertalot				+
<i>Cymbella affinis</i> Kützing		+		
<i>Cymbella cistula</i> (Ehrenberg) O. Kirchner		+	+	
<i>Cymbella leptoceros</i> (Ehrenberg) Kützing		+		+
<i>Cymbella naviculiformis</i> Auerswald ex Heiberg	+	+	+	
<i>Ecyonemasp</i>				+
<i>Eunotiabilunaris</i> (Ehrenberg) Schaarschmidt			+	
<i>Eunotia crista-galli</i> Cleve		+		
<i>Eunotia exigua</i> (Brébisson ex Kützing) Rabenhorst			+	
<i>Eunotiopaludosa</i> Grunow		+		
<i>Eunotia subarcuatoidea</i> Alles, Nörpel & Lange-Berta		+		
<i>Frustulia crassinervis</i> (Brébisson ex W. Smith) Lange-Bertalot & Krammer				+
<i>Frustulia saxonica</i> Rabenhorst			+	
<i>Gomphonema gracile</i> Ehrenberg	+	+		
<i>Gomphonema olivaceum</i> (Hornemann) Brébisson			+	
<i>Gomphonema vibrio</i> Ehrenberg			+	
<i>Hantzschia abundans</i> Lange-Bertalot	+		+	
<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	+			+
<i>Melosirasp</i>	+		+	
<i>Navicula anglica</i> Ralfs				+
<i>Navicula confervacea</i> (Kützing) Grunow				+
<i>Navicula cryptocephala</i> Kützing		+		
<i>Navicula dicephala</i> Ehrenberg			+	
<i>Navicula exigua</i> (Gregory) Grunow			+	
<i>Navicula kotschyi</i> Grunow		+		
<i>Navicula veneta</i> Kützing	+	+		
<i>Neidium amphigomphus</i> (Ehrenberg) Pfizer			+	
<i>Neidium ampliatum</i> (Ehrenberg) Krammer			+	
<i>Nitzschia denticula</i> Grunow	+	+		
<i>Nitzschia dissitata</i> (Kützing) Rabenhorst		+		
<i>Nitzschia intermedia</i> Hantzsch		+		
<i>Nitzschia linearis</i> W. Smith		+		
<i>Nitzschia palae</i> (Kützing) W. Smith				+
<i>Pinnularia anglica</i> Krammer		+		
<i>Pinnularia brauniana</i> (Grunow) Studnicka	+			
<i>Pinnularia divergens</i> W. Smith		+		

.....Continue

<i>Pinnulariamacilenta</i> Ehrenberg				
<i>Pinnularia major</i> (Kützing) Rabenhorst		+	+	
<i>Pinnulariamesolepta</i> (Ehrenberg) W. Smith	+			
<i>Pinnulariamicrostauron</i> (Ehrenberg) Cleve		+		
<i>Pinnulariaobscura</i> Krasske				+
<i>Pinnulariapseudogibba</i> Krammer		+		
<i>Pinnulariarhombarea</i> Krammer		+		
<i>Pinnulariarivularis</i>		+		
<i>Pinnulariasimilis</i> Krammer & Lange Bertalot	+			
<i>Pinnulariasubanglica</i> Krammer		+		
<i>Pinnulariasubcapitata</i> W. Gregory		+	+	
<i>Pinnulariasubstomatophora</i> Hustedt		+		
<i>Pinnulariaviridis</i> (Nitzsch) Ehrenberg	+		+	
<i>Pinnulariamesolepta</i> (Ehrenberg) W. Smith			+	
<i>Stauroneissp</i>		+		
<i>Surirellasaxonica</i> Auerswald ex Rabenhorst		+		
<i>Surirelladidyma</i> Kützing		+		
<i>Surirellalinear</i> W. Smith		+		+
<i>Surirellarobusta</i> Ehrenberg		+		+
<i>Tabellariafenestrata</i> (Lyngbye) Kützing		+		
<i>Synedraacus</i> Kützing		+		
<i>Tabellariaflocculosa</i> (Roth) Kützing		+		
<i>Tryblionellaparvula</i> (W. Smith) T. Ohtsuka & Y. Fujita			+	
Chlorophyceae				
<i>Ankistrodesmusfalcatus</i> (Corda) Ralfs		+		
<i>Asterococcusuperbus</i> (Cienkowski) Scherffel				+
<i>Bracteacoccus minor</i> (Schmidle) ex Chodat			+	
<i>Chlamydomonasdeasonii</i> Ettl		+		
<i>Chlamydomonasreinhardtii</i> P.A. Dangeard				+
<i>Chlamydomonassnowiae</i> Printz	+			
<i>Chlorococcumacidum</i> P.A. Archibald & Bold	+		+	
<i>Chlorococcumechinozygotum</i> Starr				+
<i>Chlorosarcinopsisgelatinosa</i> Chantanachat & Bold				+
<i>Desmodesmusmaximus</i> (West & G.S. West) Hegewald		+		
<i>Elliptochlorisbilobata</i> Tschermak-Woess				+
<i>Gloeocystisampla</i> (Kützing) Rabenhorst		+		
<i>Gloeocystispolydermatica</i> (Kützing) Hindak			+	
<i>Gloeocystisvesiculosa</i> Nägeli			+	
<i>Microsporaflaccida</i> (Vaucher) Thuret		+	+	
<i>Microspora pachyderma</i> (Wille) Lagerheim			+	
<i>Microspora timidula</i> Hazen	+			
<i>Neosporangiococcumgranatum</i> Deason			+	
<i>Neosporangiococcumvacuolatum</i> Deason & E.R. Cox			+	
<i>Scenedesmusabundans</i> (O. Kirchner) Chodat	+	+		
<i>Scenedesmusaculeolatus</i>		+		
<i>Scenedesmusarcuatus</i> (Lemmermann) Lemmermann		+		
<i>Scenedesmusbijuga</i> Kützing		+		+
<i>Scenedesmusdimorphus</i> (Turpin) Kützing		+		
<i>Scenedesmusflavescens</i> Chodat			+	
<i>Scenedesmus perforates</i> Lemmermann			+	
<i>Scenedesmusquadricauda</i> (Turpin) Brébisson		+		
<i>Tetrasporalacustris</i> Lemmermann	+			
Trebouxiophyceae				
<i>Chlorella ellipsoidea</i> Gerneck		+		
<i>Chlorella saccharophila</i> (Krüger) Migula			+	
<i>Chlorella vulgaris</i> Beyerrinck (Beijerinck)	+	+		+
<i>Dictyochloropsis splendens</i> Geitler				+
<i>Monoraphidium contortum</i> (Thurut) Komarkova-Legnerova	+			
<i>Monoraphidium grafitii</i> (Berkeley) Komarkova-Legnerova		+		
<i>Monoraphidium irregulare</i> (G.M. Smith) Komarkova		+		
<i>Monoraphidium minutum</i> (Nägeli) Komarkova-Legnerova			+	
<i>Monoraphidium saxatile</i> Komarkova-Legnerova	+	+		+
<i>Monoraphidium tortile</i> (West & G.S. West)			+	
<i>Murielladecolor</i> Vischer	+			
<i>Oocystis borgei</i> J. W. Snow			+	
<i>Oocystis elliptica</i> West			+	
<i>Oonophrisobesa</i> (West & G.S. West) Fott	+			
<i>Pseudococcomyxa simplex</i> (Mainx) Fott				+
<i>Stichococcus bacillaris</i> Nägeli			+	
<i>Trebouxia arboricola</i> Puymaly			+	
Ulvophyceae				
<i>Geminellamutabilis</i> (Brébisson) Wille				+
<i>Rhizoclonium</i> sp				+
<i>Ulothrix</i> sp			+	+
<i>Ulothrix tenerrima</i> (Kützing) Kützing			+	
Klebsormidiophyceae				
<i>Klebsormidium crenulatum</i> (Kützing) Lokhorst			+	
<i>Klebsormidium elegans</i> Lokhorst	+			
<i>Klebsormidium flaccidum</i> (Kützing) P.C. Silva, K.R. Mattox & W.H. Blackwell	+	+	+	+

<i>Klebsormidiumfluidans</i> (F.Gay) Lokhorst		
<i>Klebsormidiumnitens</i> (Kützing) Lokhorst		+
Zygnematophyceae		
<i>Closteriumacutum</i> Brébisson	+	
<i>Closteriumkuetzingii</i> Brébisson	+	
<i>Closteriumnavicula</i> (Brébisson) Lütkemüller		+
<i>Closteriumparvulum</i> Nägeli	+	
<i>Cosmariumabbreviacium</i> Raciborski	+	
<i>Cosmariumvariolum</i> P. Lundell	+	
Zygnemasp		+
Xanthophyceae		
<i>Tribonemaviride</i> Pascher		+
<i>Vaucheria</i> sp	+	
<i>Xanthonemaquafra</i> (Pascher) P.C. Silva		+

Table 4. Pearson's Correlation coefficient (r) values obtained between different algal class diversity (CH+X- Chlorophyta+ Xanthophyceae), CY-Cyanobacteria, D -Bacillariophyceae) with soilphysic-chemical parameters

	pH	Moisture	Conductivity	Organic carbon	Total nitrogen	Phosphorus	Potassium
Moisture	-0.52						
Conductivity	-0.60	0.63					
Organic carbon	-0.54	0.59*	0.67*				
Total nitrogen	-0.59*	0.86	0.78*	0.66*			
Phosphorus	-0.22	-0.07	0.35	0.56	0.13		
Potassium	-0.45	0.19	0.19	0.52	0.16	0.27	
CH+X	-0.51*	0.83*	0.67	0.61	0.70	-0.07	0.07
CY	-0.53	0.84*	0.68	0.62*	0.72*	-0.09	0.11
D	0.57*	0.87	0.71	0.68	0.78*	-0.02	0.14*

*Indicates significance at $p < 0.05$

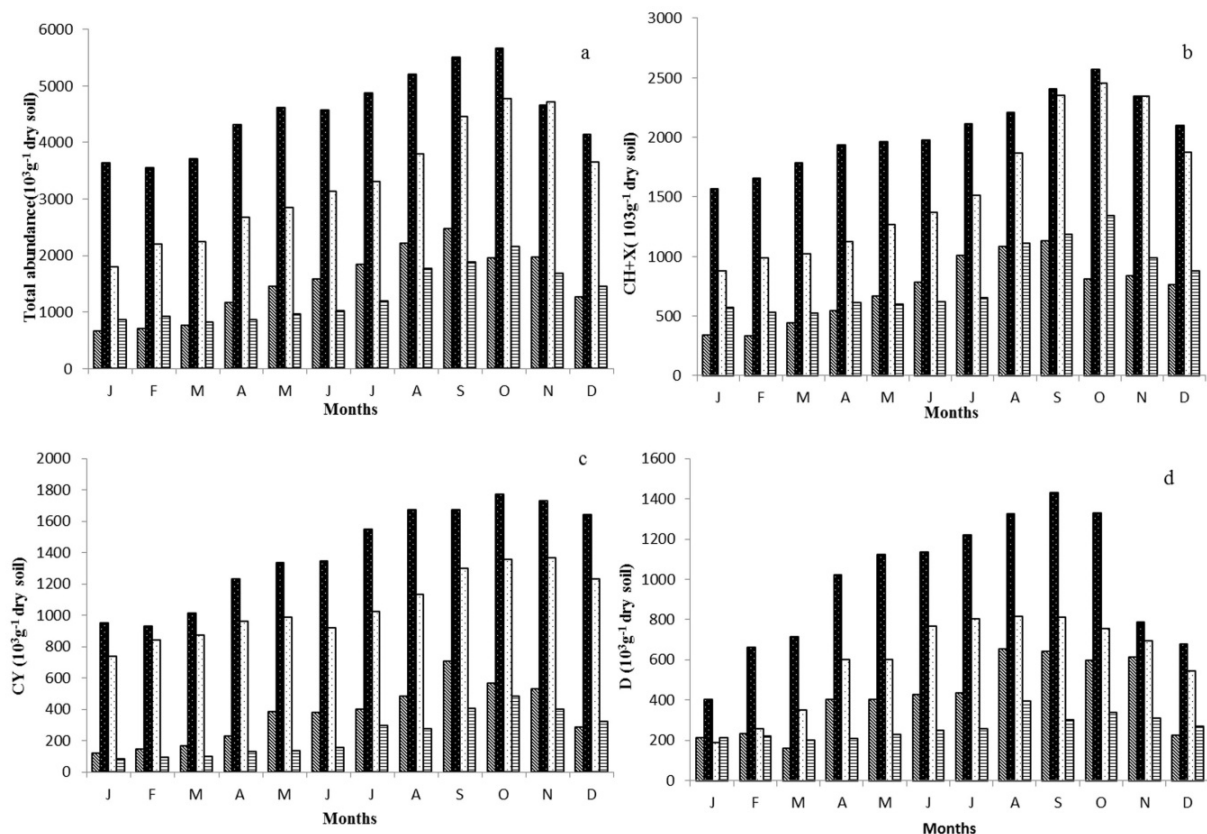


Figure 1. Algal abundance in corn field (▨), rice paddy (■), vegetable farm (▨) and citrus plantation (▨). (a). total algal abundance (b). abundance of CH+X(green algae +Xanthophyceae) (c). abundance of CY (Cyanobacteria) (d). abundance of D (Bacillariophyceae)

highest in rice field (1.55- 2.62) followed by vegetable farm (0.69-2.10) corn field (1.05-2.01) and lowest in citrus plantation (0.98-1.99) (Figure 2). The distribution of CH+ X members closely reflected the total abundance because this group accounted for high abundance (400 x 10³ to 2500 x 10³ algal cells g⁻¹ soil d.w) in all samples analysed, Cyanobacteria were more abundant in rice field (<1800 x 10³ algal cells g⁻¹ soil d.w), followed by vegetable farm (1300 x 10³ algal cells g

⁻¹ soil d.w) while abundance was low in corn field (700 x 10³ algal cells g⁻¹ soil d.w) and in citrus plantation (500 x 10³ algal cells g⁻¹ soil d.w). Diatom members were also most abundant in the rice field (1400 x 10³ algal cells g⁻¹ soil d.w) followed by vegetable farms (800 x 10³ algal cells g⁻¹ soil d.w) while it was low in corn field (700 x 10³ algal cells g⁻¹ soil d.w) and in citrus plantation (400 x 10³ algal cells g⁻¹ soil d.w) (Figure 1). The occurrence of common species varied in

different farmlands. Common species recorded in rice field were mostly Cyanobacteria members such as *Anabaena variabilis*, genera of *Oscillatoria*, along with member of diatom like *Cymbella*, *Nitzschia*, *Pinnularia*, and genera of green algae like *Scenedesmus* and *Chlorella*, while in vegetable farms Cyanobacteriagenus *Lyngbya*, genera of diatom like *Navicula* and *Hantzschia* and green filamentous genus *Microspora* were the common species.

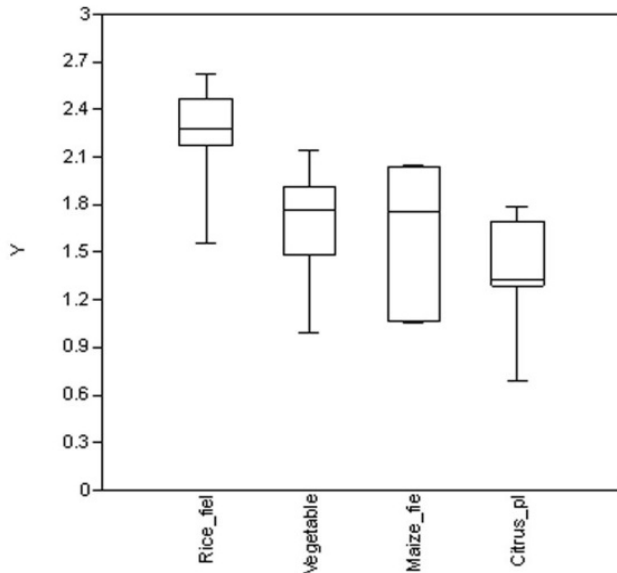


Figure 2. Boxplot of Shannon diversity index in rice field, vegetable farm, corn field and citrus plantation

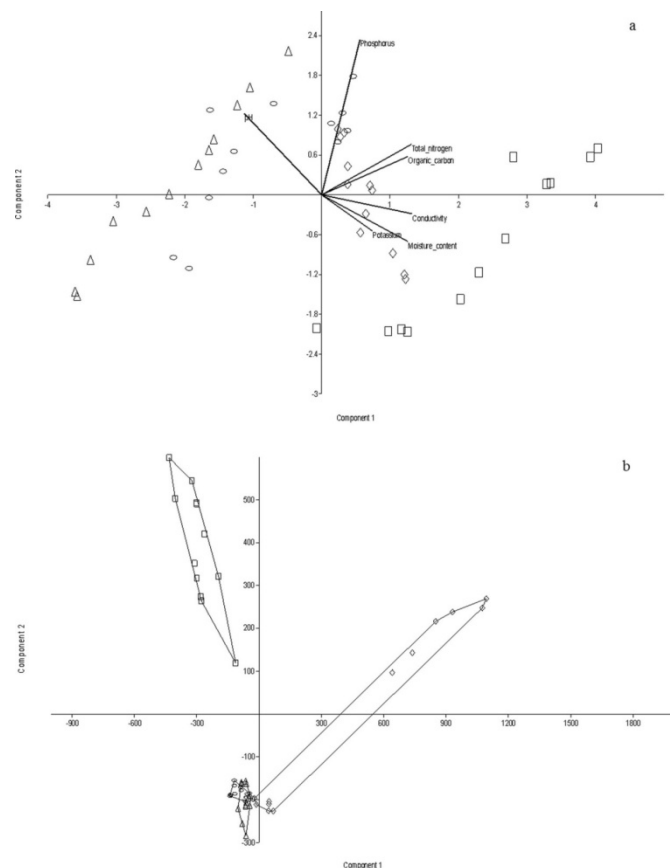


Figure 3. Ordination diagram based on (a). pH, moisture content, conductivity, organic carbon, total nitrogen, potassium and phosphorous (b). algal community variables, total algal abundance, CH+X (Chlorophyta+ Xanthophyceae), CY (Cyanobacteria) and D (Bacillariophyceae). (□) indicates rice field, (◇) vegetable farm, corn field (△) and citrus plantation (○).

The common Cyanobacteria in corn field were *Nostocmuscorum*, *Oscillatoriacurviceps*, diatom species *Hantzschiaamphioxys*, *Hantzschiaabundans* and green algal species *Chlorella vulgaris*. In citrus plantation, diatom species like *Naviculaanglica*, *Hantzschiaamphioxys* and green filamentous algae *Klebsormidiumflaccidum* and *Ulothrix* sp. were common. Correlation between CH+X diversity index with different soil parameters showed a significant positive correlation with moisture content and negative correlation with pH. Cyanobacteria diversity index showed a significant positive correlation with moisture content, carbon and nitrogen. A significant positive correlation was obtained between diversity of diatoms and pH, nitrogen and potassium (Table 4). Principal Component Analysis (PCA) was employed to compare the soil characteristics of the different farmlands, which accounted for 73% variation. Rice field could be distinguished from the other farmlands by higher moisture content, organic carbon and total nitrogen. The first principal component analysis represented 53% of the total variable, which primarily reflected the effect of moisture content, organic carbon, total nitrogen, phosphorus, potassium and electrical conductivity, while the second PC (20%) reflected the effect of pH. The first two components of the variance analysis based on algal community variables justified 64% of the variance in the data. This analysis enabled the corn field samples to be grouped together with citrus plantation samples because of the limited number of Cyanobacteria and diatoms in these farmlands. Rice field and vegetable farm are placed separately because of higher diatoms and chlorophytes. The intra-site difference was greater in vegetable farm and lower in corn field (Figure 3).

DISCUSSIONS

In the present study, pH of the soil was acidic in all the four farmlands. This is in conformity to observation by Sahu et al. (2016) that farmlands undergo intensive anthropogenic activity including tillage and application of fertilisers, pesticides and herbicides which affect the physico-chemical environment of soil and change pH to acidic condition in cultivated lands. Lin et al. (2013) also reported acidic soil condition in rice and vegetable farm in Taiwan and also related to the possible result of fertiliser application. Moisture content is an important factor in determining the composition of algal communities in all the farmlands. Dirborne and Ramanujam, (2017) reported that moisture content plays an important role in distribution and diversity of soil algae. In the present case, a comparison between soil characteristics and its relation to soil algal flora revealed highest algal diversity in rice field, which remains under high moisture content throughout the rice growing cycle, while comparatively lower algal diversity was recorded in citrus plantation with relatively low moisture content compared to other types of farmlands. Regular application of fertilisers, pesticides and insecticides in the citrus plantation, corn field and vegetable field affected the physico-chemical parameters of soil and could be responsible for low diversity of soil algae. In citrus plantation, occasional application of insecticides and pesticides were recorded to keep the trees healthy. Such activity reduced the diversity and growth of soil algae (Mostafa and Helling, 2002; Zancanet al. 2006). Application of fertilisers to enhance productivity resulted in acidic nature of the soil, which could be the reason for negative correlation of pH with nitrogen ($p < 0.05$). It is known from literature that Cyanobacteria show the most evident response to different land use patterns, and therefore considered as suitable soil bioindicator in different agro-

ecosystems. Absence of common Cyanobacteria such as *Scytonema* and rare occurrence of *Cylindrospermum* could be due to their susceptible nature to pesticides. According to Zancan *et al.* (2006) Cyanobacteria like *Scytonema* and *Cylindrospermum* occurred rarely in soil where regular application of fertilisers, pesticides and insecticides are common for better yield. Green algae like *Chlorella*, *Klebsormidium* and diatoms like *Hantzschia*, *Navicula*, *Nitzschia* and *Pinnularia* were found in all the four different farmland. All these algal species are considered as cosmopolitan and widespread in different soil types by many authors (Khaybullina *et al.*, 2010; Lin *et al.*, 2013). *Hantzschia amphioxys* and *Hantzschia abundans* diatom species are reported to have high tolerance to disturbances (Antonelli *et al.*, 2017). These two diatom species were also recorded in abundance in corn, vegetable farm and citrus plantation.

Conclusion

The structure of soil algal community is affected more by soil usage than by physico-chemical parameters of soil. The presence of adequate moisture content is one of the most important criteria for the distribution and growth of the algae. The maximum soil algal diversity was recorded in rice field where waterlogged condition is required during a part of the growing cycle. The use of fertiliser, herbicides and insecticides affected adversely the structure of algal communities.

Acknowledgement

The authors are thankful to North-eastern Hill University for providing financial assistance. Authors also acknowledge the Head of Botany Department for providing all the laboratory facilities.

REFERENCE

- Antonelli M, Wetzel CE, Ector L, Teuling AJ, Pfister L. 2017. On the potential for terrestrial diatom communities and diatom indices to identify anthropic disturbance in soils. *Ecol. Indic.* 75: 73-81.
- Automatic Diatom Identification and Classification 1999. Funded by the European MAST (Marine Science and Technology). Programme, Contract MAS3-CT97-0122.
- Desikachary, T. 1959. Cyanophyta, I. C. A. R. Monograph on algae, New Delhi.
- Dirborne CM, Ramanujam P. 2017. Diversity and ecology of soil algae in broadleaf sacred grove and pine forest in East Khasi Hills, Meghalaya. *Nelumbo* 59: 195-212
- Fritsch FE. 1935. The structure and reproduction of algae : Vol. 1 and 2 Cambridge University Press. London.
- Gandhi HP. 1998. Fresh water diatom of Central Gujarat, Bishen Sing. Mahendra Pal Singh, Shiva Offset Press
- Guiry MD, Guiry GM. 2017. AlgalBase. World-wide electronic publication, National University of Ireland, Galway.
- Hoffmann L 1989. Algae of terrestrial habitats. *Bot Rev* 55:77-105
- Hu CX, Zhang DL, Liu YD 2004. Research progress on algae of the microbial crusts in arid and semiarid regions. *Prog. Nat. Sci.* 14: 289-295
- Jackson M 1967. Soil chemical analysis, Prentice Hall Inc. Englewood Cliffs, N. J.
- John DM, Whitton BA, Brook AJ. 2002. The freshwater algal flora of the British Isles: An identification guide to freshwater and terrestrial algae, Cambridge University Press, Cambridge.
- Khaybullina LS, Gaysina LA, Johansen JR, Krautova M. 2010. Examination of the terrestrial algae of the Great Smoky Mountains National Park, USA. *Fottea* 10:201-215
- Lin C, Chou T, Wu J 2013. Biodiversity of soil algae in the farmlands of mid-Taiwan. *Bot. Stud.* 54: 41
- Lukesova A 1993. Soil algae in four secondary successional stages on abandoned fields. *Algol. Stud.* 71:81-102
- Metting, B. 1981. The systemics and ecology of soil algae. *Bot. Rev.* 47:195-312
- Mishra, U., Pabbi, S. 2004. Cyanobacteria: A potential biofertiliser for rice. *Resonance* 9:6-10
- Mostafa, F.I.Y., Helling, C.S. 2002. Impact of four pesticides on growth and metabolic activities of two photosynthetic algae. *J. Environ. Sci. Health B* 37: 417-444
- Neustupa, J. 2001. Soil algae from marlstone-substratum based biotopes in the Nature park Džbán (Central Bohemia, Czech Republic) with special attention to the natural treeless localities. *Algol. Stud.* 101:109-120
- Paoletti MG, Iovane E, Cortese M. 1988. Pedofaunabiocindicators and heavy metals in five agroecosystems in north-east Italy. *Revue D Ecologie Et De Biologie Du Sol* 25: 33-58
- Paoletti MG, Pimentel D 1992. Biodiversity in Agroecosystems. Elsevier, Amsterdam
- Philipose MT 1967. Chlorococcales, I.C.A.R., Krishi Bhawan, New Delhi.
- Prescott GW 1982. Algae of Western Great Lakes Area, Michigan State University, Otto Koelt Science Publishers, Koenigstein, West Germany.
- Ray, F.J., Thomas, B.T. 2013. Ecology and biodiversity of soil algae of Pathanamthitta District, Kerala, PhD thesis, Mahatma Gandhi University, Kerala.
- Sahu, C., Basti, S., Pradhan, R.P., Sahu, S.K. 2016. Physico-chemical properties of soil under different land use practices located near Bhawanipatna town in Odisha, India. *International Journal of Environmental sciences* 6:941-953
- Shannon, C.E., Wiener, W. 1963. The Mathematical Theory of Communication, University of Illinois Press, Urbana.
- Sukala, B.L., Davis, J.S. 1994. Algae from nonfertilized soils and soils treated with fertilizers and lime of northcentral Florida. *Nova Hedwigia* 59: 33-46
- Vijayan, D., Ray, J.D. 2015. Ecology and diversity of Cyanobacteria in Kuttanadu paddy wetlands, Kerala, *India. Am. J. Plant Sci.* 6: 2924-2938
- Walkley, A., Black, I.A. 1934. An examination of Degtjareff method for determining soil organic matter and a proposed modification of the chromic acid titration method. *Soil Sci.* 37: 29-37.
- Zancan, S., Trevisan, R., Paoletti, M.G. 2006. Soil algae composition under different agroecosystems in Northeastern Italy. *Agric. Ecosyst. Environ.* 112: 1-12.
- Zenova, G.M., Shtina, E.A., Dedysh, S.N., Glagoleva, O.B., Likhacheva, A.A., Gracheva, T.A. 1995. Ecological relations of algae in biocenoses. *Mikrobiologiya* 64: 121-133