



**Full Length Research Article**

**HISTOLOGICAL STUDIES ON THE COMBINED EFFECT OF SYNTHETIC PYRETHROID LAMBDA CYHALOTHRIN AND ORGANOPHOSPHATE CHLORPYRIFOS ON REPRODUCTIVE TISSUES IN FRESHWATER CULTIVABLE FISH *CLARIAS BATRACHUS***

\*<sup>1</sup>Saravanan, R., <sup>2</sup>Venkatesan, S. and <sup>3</sup>Revathi, K.

<sup>1,2</sup>Department of Advanced Zoology and Biotechnology, Dr Ambedkar Government Arts College, Vyasarpadi, Chennai 600039, Tamilnadu, India

<sup>3</sup>Department of Advanced Zoology and Biotechnology, Ethiraj college for women, Chennai -600008, Tamilnadu, India

**ARTICLE INFO**

**Article History:**

Received 04<sup>th</sup> March, 2015  
Received in revised form  
03<sup>rd</sup> April, 2015  
Accepted 19<sup>th</sup> May, 2015  
Published online 28<sup>th</sup> June, 2015

**Key Words:**

Lambda- cyhalothrin,  
Chlorpyrifos, Testis,  
Seminal vesicle,  
*Clarias batrachus*.

**ABSTRACT**

In the present study freshwater commercially cultivable catfish *Clarias batrachus* was used to evaluate the combined effect of lambda cyhalothrin, a synthetic pyrethroid and chlorpyrifos, an organophosphate insecticide widely used in agricultural fields. *Clarias batrachus* uses rice paddy slurries as its breeding and nursery grounds, inhabit sediments as they are benthic and are posed to greater threat from sediment associated toxic compounds such as pyrethroids and organophosphates mixtures which are widely used in rice fields today to control rice pests. The LC<sub>50</sub> of the combined mixture was found to be 0.05 mg/L. The fishes were maintained at one-fifth sublethal concentration of 0.01mg/L for a period of 30 and 45 days. Histological examination was carried out in testis and seminal vesicle at the end of exposure period to study the extent of damage at safe concentration.

Copyright © 2015 Saravanan et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**INTRODUCTION**

Residues of pesticides are found in areas far off from the point of application as they are transported into aquatic environment. The presence of pesticides within aquatic environments can induce physiological and behavioural changes in aquatic organisms although, sublethal can impair the survival and biochemical fitness of the organism (Werimo et al., 2009). Fishes raised in commercial ponds are also exposed to various pesticide mixtures from the nearby fields by runoff or spray-drift which often pollute the ponds and affect health of fish fauna (Wang et al., 2009a). Contamination of surface waters has been well documented worldwide and constitutes a major issue at local, regional, national and global levels (Glaser, 2006). The landscape in a single river catchment may have a variety of agricultural uses where a number of different crop protection programs are employed.

Agricultural practices can include pesticides being mixed during application. Therefore aquatic non-target organisms are exposed to multiple stressors including pesticides and their degradation products both simultaneously and consecutively (Colovic et al., 2011). Lambda cyhalothrin is a Type II synthetic pyrethroid and is a derivative of cyano group (Liu et al., 2004; Sudakin, 2006). Monitoring studies recently in agricultural areas show pyrethroid contamination of both surface water and stream bed sediments (Amweg et al., 2005). Investigations are ongoing on concern over bioavailability and effect on sediment and benthic dwelling organisms (Weston et al., 2004). Chlorpyrifos is a broad spectrum organophosphate insecticide which is effective in controlling insect pests (RED, 2006). Chlorpyrifos does not partition easily from soil to water, Therefore chlorpyrifos found in runoff water is likely a result of soil bound chlorpyrifos from eroding soil rather than from dissolved chlorpyrifos (Arjmandi et al., 2010). It is highly toxic to freshwater fishes and potential to bioaccumulate in the tissues as metabolites 3, 5, 6 trichloro-2-pyridinol (TCP). Sediment associated pesticides and dissolved pesticide concentration can provide a direct exposure pathway for benthic dwelling organisms and can predict a source of risk

\*Corresponding author: Saravanan, R.,

Department of Advanced Zoology and Biotechnology, Dr Ambedkar Government Arts College, Vyasarpadi, Chennai 600039, Tamilnadu, India

(Solomon et al., 2001). In the aquatic environment these hydrophobic chemical contaminants rapidly associate with particles in water and are deposited in sediments thus consumed by benthic organisms (Roessink et al., 2005). Repeated exposures and multiple stressors such as pesticides mixtures have clear relevance to real situations, it is necessary to understand their effects on non-target organisms (Ashauer et al., 2006; 2007c) so that appropriate risk assessment methodology can be developed. Zhang et al. (2010) reported effects on the 96 hr LC<sub>50</sub> in Zebrafish (*Brachydanio rerio*) exposed to binary combinations of pyrethroid insecticides permethrin, bifenthrin, tetramethrin and etofenprox with the organophosphorus insecticides dichlorvos or phoxim. The differences in the scale of interactions between pyrethroids and organophosphorus insecticides depends on both the strength of the interaction between the organophosphate and carboxylesterases and the importance of the carboxylesterase in the metabolic degradation of the pyrethroid (Groner and Relyea, 2011). The outcome of the present study would decipher the impact of low concentrations and synergistic effect of synthetic pyrethroid lambda cyhalothrin and organophosphate chlorpyrifos on the reproductive tissues. It would also ascertain whether these pesticides when present in combined doses in ambient water would have deleterious effect on various organs. The tissues chosen for the study was testis and seminal vesicle which are involved in reproductive process and play a critical role in fish breeding and development.

## MATERIALS AND METHODS

### Experimental animals

Healthy adult male catfishes *Clarias batrachus* weighing 150-175 g and 30-35 cms was used as the experimental model to evaluate the combined toxicity of lambda cyhalothrin, a synthetic pyrethroid and chlorpyrifos an organophosphate widely used for agricultural applications in various agro-based sectors. The fishes were procured from Bharath fish seed farm, Budur village, Poondi, Thiruvallur district. They were bought to the laboratory and acclimatized under laboratory conditions for a period of three weeks and fed *ad libitum*. They were transported to the laboratory in oxygen filled polythene bags and acclimatized for a period of two weeks with normal photoperiod. The fishes were maintained in rectangular plastic tubs (64cm x 44cm x 29.5cm) filled with 20 litres of dechlorinated tap water. The tubs were disinfected with potassium permanganate solution and washed thoroughly prior to introduction of fishes to prevent any fungal infection. They were fed with commercially available feed one hour prior to water exchange. Water was renewed every 24 hour to maintain healthy environment of the fish and to remove the feed remains and nitrogenous waste products. Feeding was stopped 24 hours before the commencement of the toxicity test to keep the animals more or less in the same metabolic state.

### Experimental Design

Formulated mixture of the two pesticide used in the study was supplied by Agriculture Division, Rallis India, Bangalore for evaluating the toxic effect of the pesticide mixture (EC 50) synthetic pyrethroid lambda cyhalothrin (10%) and

organophosphate chlorpyrifos (40%) on freshwater catfish *Clarias batrachus*. Preliminary toxicity tests were carried out to find the median lethal tolerance limit of experimental fishes. To determine the 96 hrs LC<sub>50</sub> static renewable bioassay method was adopted (Sprague, 1973) and 96 hr LC<sub>50</sub> was determined by probit analysis method (Finney, 1971). The LC<sub>50</sub> was found to be 0.05 mg/L. The fishes were exposed for a period of 30 and 45 days at one fifth of the sublethal concentration (0.01 mg/L). The fishes were divided into 2 groups with five fishes in each group. Group I: Control fishes in toxicant-free water, Group II: Fishes maintained in 0.01 mg/L of pesticide mixture for a period of 30 days. Group III: Fishes maintained in 0.01 mg/L of pesticide mixture for a period of 45 days.

### Collection of tissues for histological studies

The animals were sacrificed, testis and seminal vesicle was separated and immediately fixed in Bouin's fluid for histological studies following the method of Roberts (1978) and Humanson (1979). The sections were stained with haematoxylin and eosin.

## RESULTS

Histological studies of the reproductive tissues showed marked and profound changes. The following changes were noticed in the testis and seminal vesicle of *Clarias batrachus* exposed to the pesticidal combination during 30<sup>th</sup> and 45<sup>th</sup> day of exposure.

**Testis:** Normal testis showing seminiferous tubules containing cells at different spermatogenic stages (Fig 1a). During 30 days of exposure to the pesticidal mixture extensive cytotoxic damage with large number of vacuoles were prominent. Gross condensation of spermatogenic cells. Severe degeneration and displacement of tubular elements (Fig 1b). Distortion of tubular elements increased with the number of days of exposure. Appearance of vacuoles in the tubules. Seminiferous tubules of central region show degeneration and reduction with disappearance of spermatozoa and spermatogenic cells were evident on the 45<sup>th</sup> day of exposure (Fig 1c).

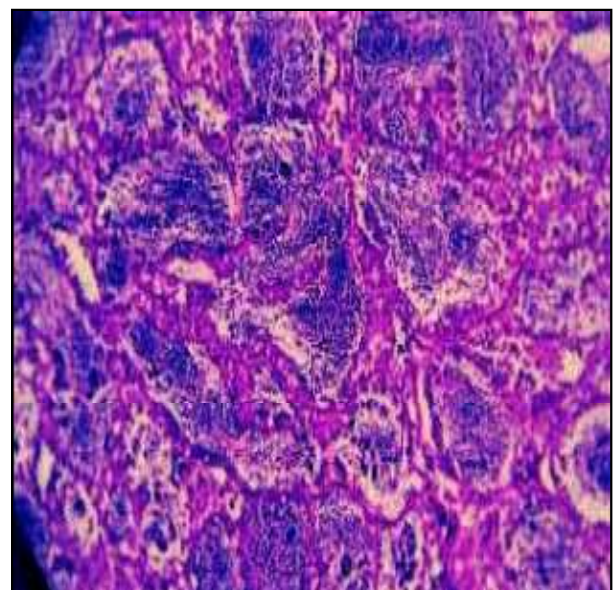


Fig. 1a. Normal Testis of *Clarias batrachus* (100x)

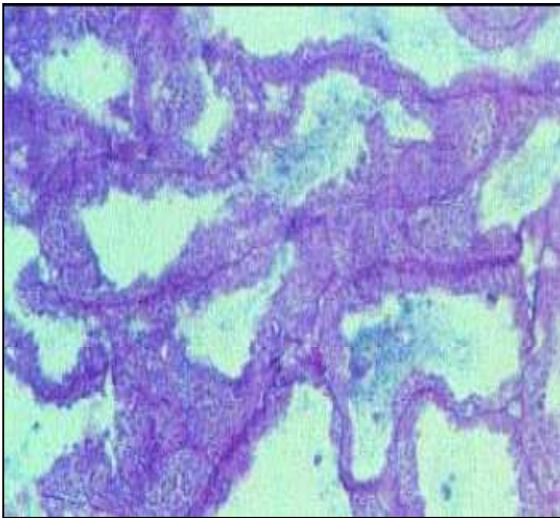


Fig. 1b. Shows vacuolization and degeneration of seminiferous tubules of testis (arrows) during 30 days of exposure (100x)

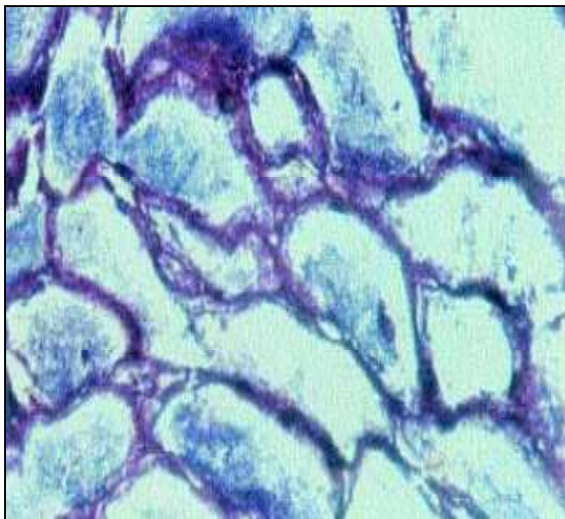


Fig. 1c. Shows vacuolization and degeneration of seminiferous tubules of testis (arrows) during 30 days of exposure (100x)

**Seminal vesicle:** Normal seminal vesicle showing growth buds, epithelium, new lobules and spermatozoa in the lumen (Fig 2a).

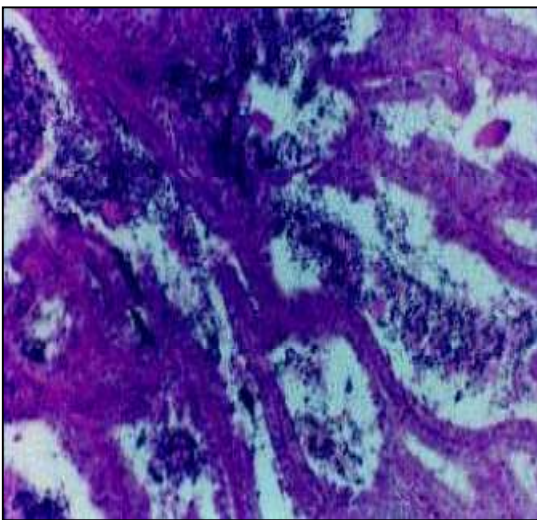


Fig. 2a. Normal Seminal vesicle of *Clarias batrachus* (100x)

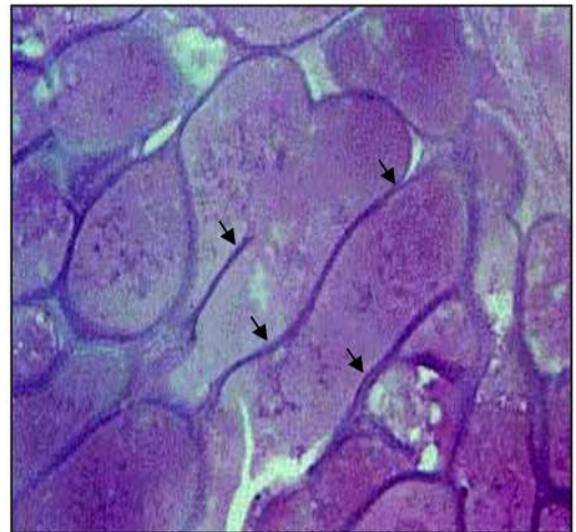


Fig. 2b. Shows epithelial cell lining damage and contents of lumen empty (arrows) during 30 days of exposure (100x)

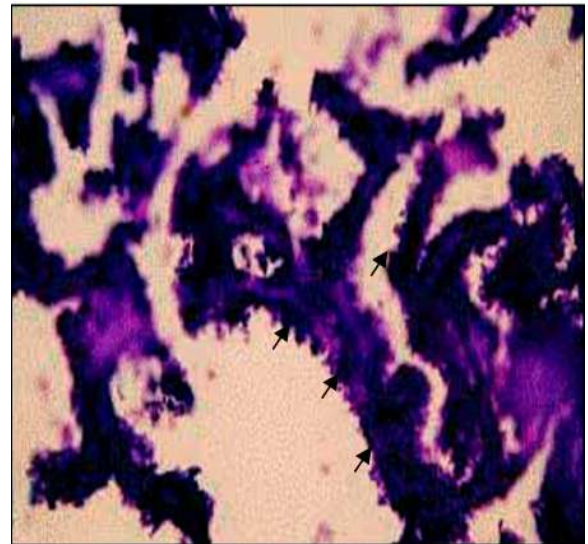


Fig. 2c. Shows epithelial cells desquamation and lumen lobules degeneration (arrows) during 45 days of exposure (100x)

Lobulated lumen empty, collapsed epithelium of lumen during 30 days of exposure (Fig 2b), epithelium lining damaged, absence of vesicular contents and lobes of seminal vesicle could be observed with disorientation during 45<sup>th</sup> day of exposure (Fig 2c).

## DISCUSSION

In the fishes exposed to lambda cyhalothrin and chlorpyrifos mixture seminiferous tubule vacuolization was observed with decreased number of developing spermatocyte of different stages in the testis. Colloidal mass was observed in the vacuoles. After 45 days of exposure there was central region degeneration of testis, shrinkage of interstitial cells with reduced number of developing germ cells. The effect of lambda cyhalothrin could have brought about retardation of testicular functions and degeneration of testis with seminiferous tubules losing their shape. The spermatogonia and spermatocytes lose their characteristics and are greatly reduced in number. Kinneberg

et al. (2000) have also documented concentration dependent effects on nonylphenol on testicular structure of the fish *Xiphophorus maculatus*. *Glossogobius giuris* exposed to fenthion showed reduction in size of testis, degenerating sperms and spermatids (Zutschi, 2003). Direct toxic effects of diazinon was noticed in cyprinid fish *Rutilus kutum* with degeneration of seminiferous tubules and low quality of sperm (Fadakar Masouleh, 2011), testis of blue gill fish (Dutta and Meijer, 2003; Maxwell and Dutta, 2005) and common carp (Banaee et al., 2009). Disorganized testicular architecture and disrupted germinal epithelium was reported in air-breathing catfish *Heteropneustes fossilis* exposed to cadmium (Shivani Sharma et al., 2013). Impairment in reproduction have been observed with a number of pollutants in freshwater and marine fishes (Govind and Madhuri, 2014). The seminal vesicle fluid is secreted by the epithelium lining the tubules.

The tubules in the proximal part of the lobes are predominantly lined by a simple cylindrical and those of the distal part by a simple squamous epithelium. These epithelial cells contain enzymes involved in energy-liberating processes, the enzyme activities being proportional to the height of the cells (Fishelson et al., 1994). The lobulated lumen showed less luminal contents on the 30<sup>th</sup> day with epithelial lining damage. The complete epithelium was disoriented and lobule damage of the seminal vesicle was evident in the fishes exposed to the pesticidal mixture for 45 days.

The study suggest that reproductive tissues are susceptible to high degree of damage even at very low toxicant of different combinations present in ambient water. Moreover, considering the commercial importance of this freshwater fish, the present study has implication for the aquaculture industry which may lead to reduction in maturation associated growth, reproductive vigour and deterioration in flesh quality leading to heavy loss in fish production.

## REFERENCES

Amweg, E.L., Weston, D.P. and Ureda, N.M. 2005. Use and toxicity of pyrethroid pesticides in the Central Valley, California, USA. *Env. Toxicol. Chem.*, 24: 966.

Arjmandi, R., Tavakol, M. and Shayeghi, M. 2010. Determination of organophosphorus insect residues in the rice paddies. *Int. J. Env. Sci. Technol.*, 7 (1): 175-182

Ashauer, R., Boxall, A. B. A. and Brown, C. D. 2006. Predicting effects on aquatic organisms from fluctuating or pulsed exposure to pesticides. *Environ. Toxicol. Chem.*, 25 (11): 1899.

Ashauer, R., Boxall, A. B. A. and Brown, C. D. 2007c. Modeling combined effects of pulsed exposure to carbaryl and chlorpyrifos on *Gammarus pulex*. *Environ. Sci. Technol.*, 41 (15): 5535-5541.

Banaee, M., Mirvaghefi, A.R., Ahmadi, K. and Ashori, R. 2009. The effect of diazinon on histopathological changes in testis and ovaries of common carp (*Cyprinus carpio*). *Sci. J. Mar. Biol.*, 1(2): 25-35.

Colovic, M. B., Krstic, D. Z., Uscumlic, G.S. and Vasic, V. M. 2011. Single and simultaneous exposure of acetyl cholinesterase to diazinon, chlorpyrifos and their photo degradation products. *Pest. Biochem. Physiol.*, 100(1): 16-22.

Dutta, H.M. and Meijer, H.J.M. 2003. Sublethal effects of diazinon on the structure of testis of bluegill *Lepomis macrochirus*: a microscopic analysis. *Env. Poll.*, 125: 355-360

Fadakar Masouleh, F. Mojazi Amiri, B., Mirvaghefi, A.R. and Nematollahi, M. A. 2011. *In vitro* effects of diazinon on male reproductive tissue and sperm motility of Caspian Kutum (*Rutilus Frisii Kutum*). *Res. J. Env. Toxicol.*, 5(2): 108-116.

Finney, D.J. 1971. Probit analysis-A statistical treatment of sigmoid curve-(3<sup>rd</sup> edn). Cambridge University Press, London. p 568.

Fishelson, L., Van Vuren, J.H.J. and Tyran, A. 1994. Ontogenesis and ultrastructure of seminal vesicles of catfish *Clarias gariepinus*. *J. Morphol.*, 219: 59-71

Glaser, A. 2006. Threatened waters. Turning the tide on pesticide contamination. *Beyond pesticides*, 1(2) : 2-5 .

Govind, P and Madhuri, S. 2014. Heavy metals causing toxicity in animals and plants. *Res J Animal, Veterinary and Fishery Sci* ,2 (2), 17-21.

Groner, M.L. and Relyea, R.A. 2011. A tale of two pesticides: How common insecticides affect aquatic communities. *Freshwater. Biol.*, 6(11): 2391-2404.

Humanson, G. L. 1979. *Animal tissue technique* (4<sup>th</sup> ed). W. H Freeman and Company, San Francisco, USA. p 3-63.

Kinnberg, K., Korsgaard, B. and Bjerregaard, P. 2000. Concentration dependent effects of nonylphenol on testis structure in adult platyfish, *Xiphophorus maculatus*. *Mar. Environ. Res.*, 50: 169-173

Liu, W., Gan, J. J, Lee, S. and Kabashima, J.N. 2004. Phase distribution of synthetic pyrethroid in runoff and stream waters. *Environ. Toxicol. Chem.*, 23: 7.

Maxwell, L.B. and Dutta, H.M. 2005. Diazinon induced endocrine disruption in bluegill sunfish *Lepomis macrochirus*. *Ecotoxicol. Environ. Safety.*, 60:21-27

Shivani Sharma, Vipin Vyas, Sadhna Tamot and Susan Manhor, 2014. Histological changes in the testis of air-breathing fish, *Heteropneustes fossilis* following cadmium chloride exposure. *J Chem Bio Phy Sci*, 3 (2):1216-1221.

Solomon, K. R, Giddings, J.M. and Maund, S.J. 2001. Probabilistic risk assessment of cotton pyrethroids. Distributional analysis of laboratory aquatic toxicity data.. *Env. Toxicol Chem.*, 20: 652

Sprague, J.B. 1973. Measurement of pollutant toxicology of fish. Sublethal effects and safe concentration. *Water Res*, 5: 245-266.

Sudakin, D.L. 2006. Pyrethroid insecticides: advances and challenges in biomonitoring. *Clin Toxicol*, 44: 31-39

Reregistration Eligibility Decision (RED) for Chlorpyrifos, 2006. U.S. Environmental Protection Agency, Office of Prevention, Pesticides and Toxic Substances, Office of Pesticide Programs, U.S. Government Printing Office: Washington, DC.

Roberts, R. J. 1978. The pathophysiology and systemic pathology of teleosts and laboratory methods. In: *Fish Pathology* (1<sup>st</sup> ed). Bailliere Tindall London, UK, p 235-246

Roessink, I. G., Arts, H. P., Belgers, J.D.M., Bransen, F., Maund, S.J. and Brock, T.C.M. 2005. Effects of lambda cyhalothrin in two ditch microcosm systems of different trophic status. *Env. Toxicol. Chem.*, 24: 1684 -1690

- Wang, C., Lu, G., Cui, J. and Wang, P. 2009a. Sublethal effects of pesticide mixtures on selected biomarkers of *Carassius auratus*. *Envt. Toxicol. Pharmacol.*, 28(3): 414-419.
- Werimo, K., Bergwerff, A.A. and Seinen, W. 2009. Residue levels of organochlorines and organophosphates in water, fish and sediments from Lake Victoria Kenyan portion. *Aquat Eco. Health. Mgmt.*, 12: 337-341.
- Weston, D. P., You, J. and Lydy M. J. 2004. Distribution and toxicity of sediment associated pesticides in agricultural dominated water bodies of California's Central Valley. *Envt. Sci. Tech.*, 38: 2752.
- Zhang, Y., Liu, S., Liu, H. and Liu, Z. 2010. Evaluation of the combined toxicity of 15 pesticides by uniform design. *Pest. Mangt. Sci.*, 66(8): 879-887.
- Zutshi, B. 2005. Ultrastructural studies on the effect of fenthion on pituitary (GTH cells) and testis of *Glossogobius giuris* (Ham) during breeding phase. *J. Environ. Biol.*, 26: 31-36.

\*\*\*\*\*