



Full Length Research Article

ANTIBACTERIAL ACTIVITY OF *EXCOECARIA AGALLOCHA* L. LEAF IN CHLOROFORM AND ETHANOL EXTRACTS (GC-MS ANALYSIS)

¹Parasuraman, P. and ²Dr. Manikandan, T.

¹Govt. Hr. Secodary School, Devikapuram, Tiruvannamalai-606902, Tamilnadu, India

²Department of Botany, Arignar Anna Govt. Arts College, Villupuram- 605602, Tamilnadu, India

ARTICLE INFO

Article History:

Received 16th November, 2016
Received in revised form
24th December, 2016
Accepted 21st January, 2017
Published online 28th February, 2017

Key Words:

Excoecaria agallocha L.
Antibacterial Activity,
Chloroform, Ethanol,
GC-MS Analysis.

ABSTRACT

The present study was carried out for Antibacterial Activity and GC-MS analysis of *Excoecaria agallocha* L. leaf extracts of Chloroform and Ethanol. Antibacterial assay was carried out against five bacteria viz. *Staphylococcus aureus* MTCC3381, *Bacillus cereus* MTCC430, *Escherichia coli* MTCC739, *Pseudomonas aeruginosa* MTCC 424, *Klebsiella pneumoniae* MTCC432 using agar well diffusion method. Chloroform extract of *E.agallocha* was highly effective on *Bacillus cereus* strain. There was no effect on *E.coli* and *K.pneumoniae*. It was moderately effective in 2000µg concentration only. *P.aeruginosa* was affected moderately in 1500 and 2000µg concentrations. Ethanol extract was highly effective than Chloroform extract. Ethanol extract was ineffective in 500µg on *E.coli*, *Bacillus cereus*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae*. In 1500µg Ethanol extract was ineffective on *P.aeruginosa*. Ethanol extract was ineffective in 1000µg concentration on *K.pneumoniae*. It was highly effective on *S.aureus* at 500,1000,1500,2000µgs. GC-MS analysis was tested in chloroform and ethanol extracts of *E.agallocha* L. In Chloroform extract 34 bio-active compounds were identified. From that Hentriacontane 5.23%, Tricyclo, undec-tetramethyl 16.38%, Tetramethyl, trihydro-naphthalene 20.72%, β - amyryl 7.52%, α - amyryl 6.92%, Heptadecanol 8.24% were in high proportions. Ethanol extract showed 17 bio-active compounds, in that Myoinositol 4-c-methyl 37.09%, *Tricyclo undec* (isocaryophyllene) 13.21%, Trimethyl 5,6-dimethylene-deca hydro naphthalene 17.25%, β- amyryl 5.78%, α- amyryl 5.78% were observed as the major constituents.

Copyright©2016, Parasuraman and Dr. Manikandan. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Indian medicinal plants having therapeutic values and can also be used in drugs. Most of the people in developing countries depend on traditional medicines from plants for the primary health care needs as estimated by WHO. Plant medicines have minimal toxicity, low cost, pharmacologically active and provide easy remedy for human beings. The effectiveness of plant extracts on micro organisms has been studied worldwide. *Excoecaria agallocha* L. belongs to Euphobiaceae family. It is a small mangrove tree with 15m height. The bark oil is effective against rheumatism, leprosy and paralysis. However it cause temporary blindness if it enters the eyes. The plant is potent as anti- HIV, anti- cancer, anti-bacterial and anti-viral agent (Peter *et al.*, 1999). The plant is used as fire wood, timber and also gives tannin, fish poison and medicines

for epilepsy, ulcers, hand and feet swellings, toothache (Bandara Nayake, 2002). People in coastal areas destroying the tree for fodder, fuel, wood pulp vegetable tannins, poles for building constructions and medicines (UNEP 1988). Medicines from mangrove plants are used for curing elephantiasis and abdominal troubles. The main aim of the present study is the antibacterial activity and bio-active compounds of *E.agallocha* L. leaf extract in different solvents i.e., Chloroform and Ethanol.

MATERIALS AND METHODS

Mature leaves of *Excoecaria agallocha* L. were collected during monsoon period in the mangrove belt of Pichavaram, Tami Nadu, India. The present investigation was carried out in the Botanical Garden of Arignar Anna Govt. Arts College, Villupuram, Tamil Nadu, India. The plant material was identified with the help of "FLORA OF THE PRESIDENCY OF MADRAS" (Gamble, 1954). Five different bacteria were used for this study. They were *Staphylococcus aureus*

*Corresponding author: Dr. Manikandan, T.

Department of Botany, Arignar Anna Govt. Arts College, Villupuram- 605602, Tamilnadu, India.

MTCC3381, *Bacillus cereus* MTCC430, *Escherichia coli* MTCC739, *Pseudomonas aeruginosa* MTCC 424, *Klebsiella pneumoniae* MTCC432 from that *S.aureus* and *B.cereus* are gram positive strains. Rest of the three are gram negative strains (Christian Gram, 1884).

Preparation of plant extracts

For the preparation of leaf extracts Soxhlet extractor (Franz von Soxhlet in 1879) was used. The leaves were washed in tap water, shade dried and made into a fine powder. The powder was extracted in the thimble of Soxhlet extractor with the solvents Chloroform and Ethanol successively. This extracts were concentrated to dryness using rotary vacuum evaporator.

Test Organisms

The extract was tested on the following two gram positive bacteria namely *S.aureus* MTCC3381, *B.cereus* MTCC430. Three gram negative bacteria were also tested. They are *E.coli* MTCC739, *P.aeruginosa* MTCC424, *K.pneumoniae* MTCC432. All the strains were collected from Manian Institute of Science and Technology, Coimbatore, Tamil Nadu, India. Test was conducted with the help of MISAT Lab Coimbatore, Tamil Nadu.

Preparation of Inocula

Inoculation was done with the help of MISAT lab Coimbatore. The test organisms were sub cultured by streaking them on nutrient agar followed by incubation for 24h at 37°C. Several colonies of each bacteria species were transferred to sterile nutrient broth. The suspensions were mixed for 15sec and incubate for 24h at 37°C on an orbital incubated shaker. Working concentrations of microbial suspension was prepared in 3ml of sterile saline to turbidity equivalent to 0.5mc land scale (10 adjusting the potical density to 0.1 at 600nm yielding a cell density of $1-2 \times 10^5$ CFU/ml.).

GC-MS Analysis

Chloroform and Ethanol extracts were prepared with the help of Bureau Veritas Consumer product Services (I) Pvt. Ltd. Chennai-32 by GC-MS 5975C Agilent instrument. 100gms of powder was weighed and extracted using 100ml of solvents successively with Chloroform and Ethanol with the help of Soxhlet apparatus. The extracts were evaporated to dryness using rotary evaporator. The dried extract was then subjected to GC-MS Analysis. Gas chromatography technique separates chemicals based on their volatility or ease with which they evaporate into gas. The MS is used to identify the chemicals based on their structure. One micro liter (1 μ l or 0.000001L) of solvents containing the mixture of molecules were injected into the GC and the sample was carried by Helium through the instrument. Chemicals with high volatility travel through the column more quickly than chemicals with low volatility. The ions travelled through an electromagnetic field then filtrate the ion based on their mass. The spectrum of unknown components was compared with spectrum of the known components stored in NIST library. The name, molecular weight and structure of the components of the test materials were determined.

RESULTS AND DISCUSSION

Chloroform extract of *E.agallocha* showed mild effect on the micro organisms. The extract showed no inhibition on *E.coli* and *K. pneumonia* (Gram negative strains). *B.cereus* was inhibited in the range of 10.0 ± 0.0 , 11.5 ± 0.7 , 13.0 ± 0.0 , 15.0 ± 0.0 depends on the concentrations.(500,1000,1500 and 2000 μ gs). *Staphylococcus aureus* was affected only in the 2000 μ g conc.(10.0 ± 0.0). *P.aereus* was affected in 1500 μ g (10.0 ± 0.0) and in 2000 μ g (10.5 ± 0.7) (Table 1 and Plate 1). The inhibitory effect of Chloroform and Ethanol extracts of *Excoecaria agallocha* L. increases with increase in concentration. Chloroform extracts of *Solanum trilobatum* showsinhibition on *S.aureus*, *E.coli*, *K.pneumoniae*, *P.aeruginosa*. This is a negative result with *E.agallocha* L.

Table 1. Antibacterial activity of *Excoecaria agallocha* L. leaf in chloroform extract

Sample	Conc.(μ g)	Zone of Inhibition(mm)				
		S.a	E.c	B.c	P.a	K.p
Chloroform	500	-	-	10.0 ± 0.0	-	-
	1000	-	-	11.5 ± 0.7	-	-
	1500	-	-	13.0 ± 0.0	10.0 ± 0.0	-
	2000	10.0 ± 0.0	-	15.0 ± 0.0	10.5 ± 0.7	-
Chloromphenicol	10	20.5 ± 0.7	14.0 ± 0.0	20.5 ± 0.7	14.0 ± 0.0	19.5 ± 0.7

Values are means of three independent analysis \pm Standard Deviation (n=3)

Antibacterial Activity

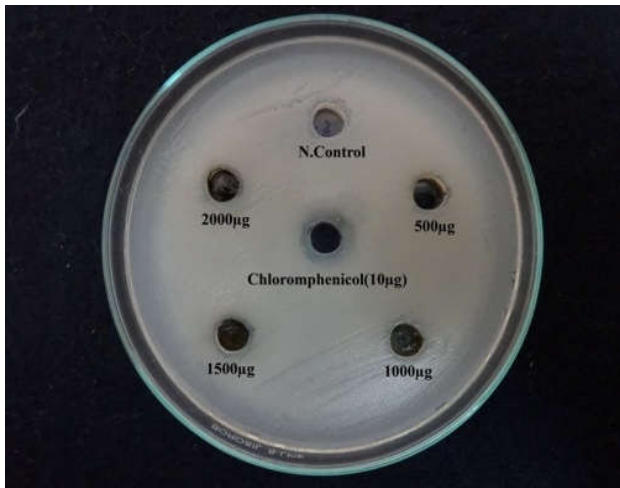
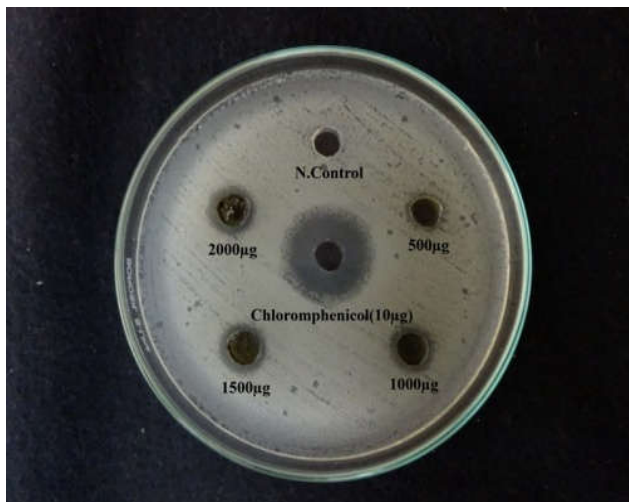
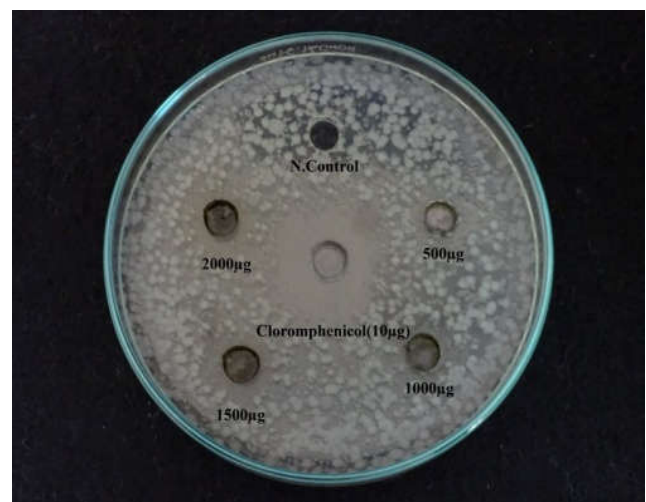
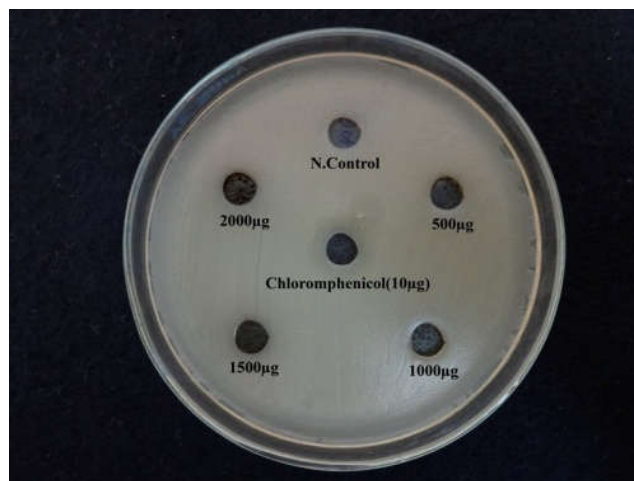
Nutrient agar (NA) plates (12cm diameter) were seeded with 8h broth culture of different bacteria. In each of this plates well were (6mm diameter) cut out using sterile cork borer. Using sterilized dropping pipettes different concentration (500,1000, 1500 and 2000 μ g /ml) of plant extract was carefully added in the wells and allowed to diffuse at room temperature for 2h (well diffusion Method Perez *et al.*,1990). The plates were then incubated at 37°C for 18-24h. Chloromphenicol (10 μ g) was used as positive control and DMSO (Dimethyl sulphoxide) as negative control. The antimicrobial activity was evaluated by measuring the diameter of inhibition zone (Agwa *et al.*, 2000). Antibacterial activity was assigned by measuring the inhibition zone formed around the discs.

leaf extract (Asirvatham doss and Rangaswamy Dhanabalan, 2008). The Chloroform extract of *E.agallocha* inhibits (50mg/ml) on *P.aeruginosa*, *S.aureus* and *E.coli* (Jayanta Kumar Patra *et al.*, 2009). But this is not similar for this present study. *Calotropis gigantia* Linn. Leaf extract of chloroform shows no inhibition on *S.aureus* and *E.coli*. It is similar to *E.agallocha* L. (Bharathi *et al.*, 2011). In 100mg/ml Chloroform extract of *Solanum nigrum* Linn. Leaf shows no effect on *E.coli* and *S.aureus* but effective on *K.pneumoniae* (10mm diameter, Sridhar *et al.*, 2011). It is similar with the present study. The Chloroform extract of *Casuarina equisetifolia* shows inhibition in 100mg/ml on *S.aureus*, *E.coli*, *K.pneumoniae* and *P.aeruginosa* (Nehad Gungumjee *et al.*, 2012). Chloroform extract of *Parthenium hysterophorus* leaf shows 63% inhibition on *B.cereus* (Malarkodi and Manoharan, 2013). It is similar with the present study.

Table 2. Antibacterial activity of *Excoecaria agallocha* L. leaf in ethanol extract

Sample	Conc (μg)	Zone of Inhibition (mm)				
		S.a	E.c	B.c	P.a	K.p
Ethanol	500	18.0 \pm 0.0	-	-	-	-
	1000	20.5 \pm 0.7	10.0 \pm 0.7	11.0 \pm 0.0	-	-
	1500	22.0 \pm 0.0	11.0 \pm 0.0	13.5 \pm 0.7	-	11.0 \pm 0.0
	2000	24.5 \pm 0.7	13.0 \pm 0.0	15.0 \pm 0.0	13.0 \pm 0.0	10.0 \pm 0.0
Chloromphenicol	10	20.5 \pm 0.7	14.0 \pm 0.0	20.5 \pm 0.7	14.0 \pm 0.0	19.5 \pm 0.7

Values are means of three independent analysis \pm Standard Deviation(n=3)

Plate 1. Antibacterial activity of *Excoecaria agallocha* L. leaf in Chloroform extract*Excoecaria Agallocha* Chloroform (S.a)*Excoecaria Agallocha* Chloroform (E.c)*Excoecaria Agallocha* Chloroform (B.c)*Excoecaria Agallocha* Chloroform (P.a)*Excoecaria Agallocha* Chloroform (K.p)

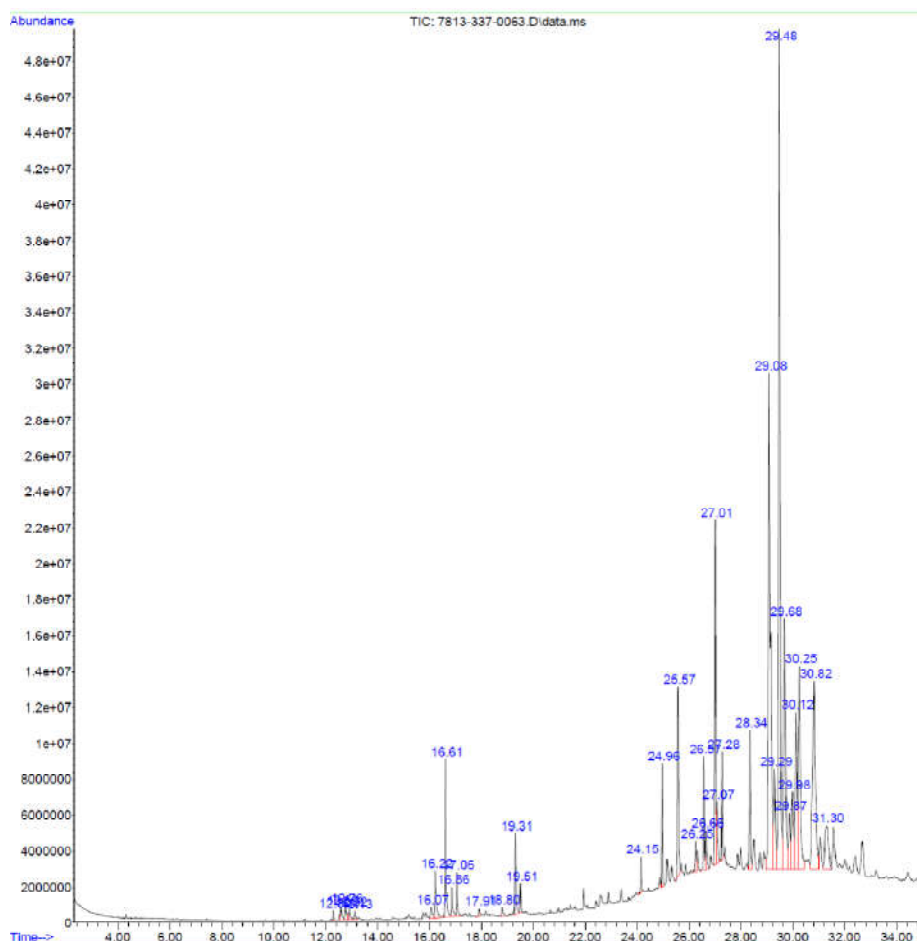


Fig.1. Peak area of Bio-active Chemicals in Chloroform Extract of Excoecaria agallocha L. leaf (GC-MS)

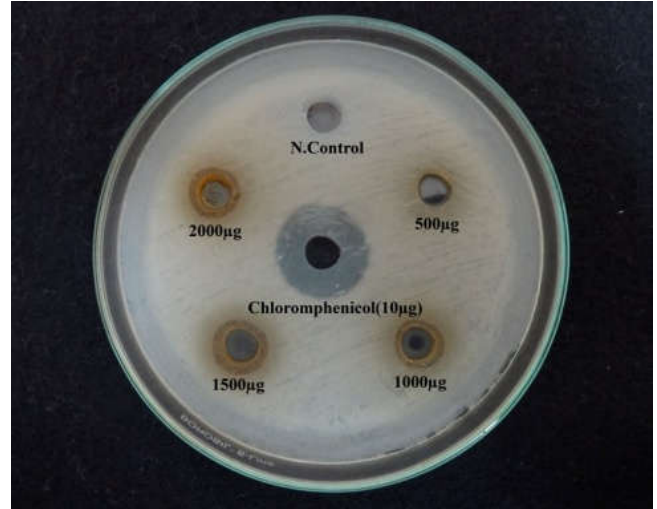
Table 3. Bio-active chemicals identified in Excoecariaagallocha L. (Chloroform extract) (GC-MS Report TLC)

Peak	R.Time(Mins)	Name of the Chemical Compound	M.F.	M.W.	Peak Area	Area%	
1	12.293	B-Curcumene	C ₁₅ H ₂₄	204	12735905	0.13	
2	12.598	Benzene ,1-(1,5 dimethyl-4-Hexenyl-4-Methyl-	C ₁₅ H ₂₂	202	26819666	0.28	
3	12.758	1,3-Cyclohexadiene,5(1,5 dimethyl)	C ₁₅ H ₂₄	204	26126978	0.28	
4	12.918	β-Bisabolene	C ₆ H ₈	080	22936539	0.24	
5	13.135	Cyclohexane,3(1,5 Dimethyl-4-Hexenyl -6-Methylene-s-(RS)	C ₁₅ H ₂₄	204	14225398	0.15	
6	16.069	5-Ethylcyclopent-1-ene-1-Carboxylic acid	C ₈ H ₁₂ O ₂	140	32786260	0.35	
7	16.229	2-Cyclohexane-1-one 4-Hydroxy- 3,5,6-Trimethyl-4(3-oxo-1Butenyl)	C ₁₃ H ₁₈ O ₃	222	107623763	1.14	
8	16.606	Bicyclo(3,1.1)Heptane,2,6,6 Trimethyl-	(C ₁₀ H ₁₆) _n	000	230080098	2.43	
9	16.868	Phytol, Acitrate	C ₂₂ H ₄₂ O ₂	338	59574193	0.54	
10	17.057	1.Hexadecyne	C ₁₆ H ₃₂	224	67813854	0.72	
11	17.914	n-Hexadecanoic acid(palmitic acid)	C ₁₆ H ₃₂ O ₂	256	11201392	0.12	
12	18.799	1H-Indazole,5,7 Dimethyl	C ₉ H ₁₀ N ₂	146	18080641	0.19	
13	19.308	Phytol	C ₂₀ H ₄₀ O	296	124447096	1.31	
14	19.511	5-Methoxy-2-Methylindole-3-acetic acid -Tertbutyl ester	C ₁₂ H ₁₃ NO ₃	219	31102439	0.33	
15	24.144	Eicosane	C ₂₀ H ₃₈	278	46352703	0.49	
16	24.957	Squalene	C ₃₀ H ₅₀	410	128286858	1.35	
17	25.576	Heptadecane,9-octyl	C ₂₅ H ₅₂	352	277558171	2.93	
18	26.250	Heptadecane	C ₁₇ H ₃₆	240	54645644	0.58	
19	26.569	1,19-Eicosadiene	C ₂₀ H ₃₈	278	146660431	1.55	
20	26.656	Gamma-Tocopherol	C ₂₈ H ₄₈ O ₂	416	71959973	0.76	
21	27.005	Hentriacontane	C ₃₁ H ₆₄	436	495524359	5.23	
22	27.063	1-Heptacosanol	C ₂₇ H ₅₄ O	394	105516830	1.11	
23	27.281	Vitamin E	C ₂₉ H ₅₀ O ₂	430	153143961	1.62	
24	28.341	Oxirane,Hexadecyl	C ₁₅ H ₃₂ O	240	251390498	2.66	
25	29.082	Tricyclo(6.2.1.0(4,11)Undecene 1,5,9,9- Tetramethyl	C ₁₅ H ₂₄	204	1551277372	16.38	
26	29.285	Androstan-17-one 16,16-dimethyl-(5-α)	C ₂₂ H ₃₆ O ₂	332	262470321	2.77	
27	29.474	2,5,5,8a-Tetramethyl-6,7-8,8a- Tetrahydro-5H-Naphthalene-1-one	C ₁₀ H ₁₈	138	1961317086	20.72	
28	29.677	β-Amyrin	C ₃₀ H ₅₀ O	426	712193762	7.52	
29	29.866	Caparratriene	C ₁₅ H ₂₆	206	157222150	1.66	
30	29.982	1,2,5-Oxadiazol-3-amine,4-(3- Methoxy Phenoxy)-	C ₁₇ H ₁₃ F ₃ N ₆ O	000	218568259	2.31	
31	30.127	9,19-Cyclo lanost-24-en-3-ol,(3β)	C ₃₀ H ₅₀	426	422863456	4.47	
32	30.244	α-Amyrin	C ₃₀ H ₅₀ O	426	655423785	6.92	
33	30.825	Heptadecanal	C ₁₇ H ₃₄ O	254	780119808	8.24	
34	31.289	2(Acetoxy methyl)3-Methoxy Carbonyl) bi phenylene	C ₁₇ H ₁₄ O ₄	282	237767401	2.51	
					Total	7697818050	99.99

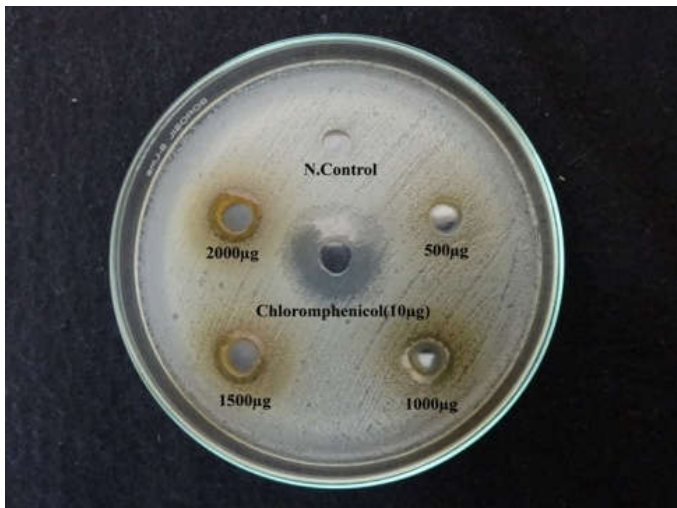
Plate 2. Antibacterial activity of *Excoecaria agallocha* L. leaf in Ethanol Extract



Excoecaria Agallocha Ethanol (S.a)



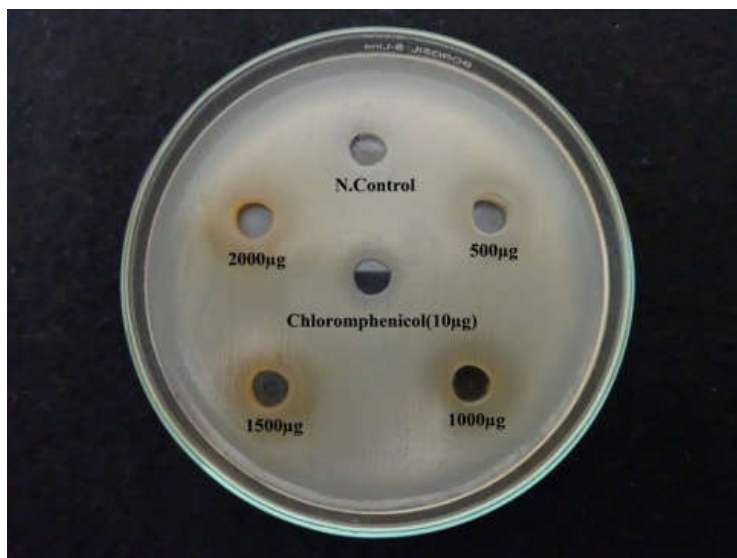
Excoecaria Agallocha Ethanol (E.c)



Excoecaria Agallocha Ethanol (B.c)



Excoecaria Agallocha Ethanol (P.a)



Excoecaria Agallocha Ethanol (K.p)

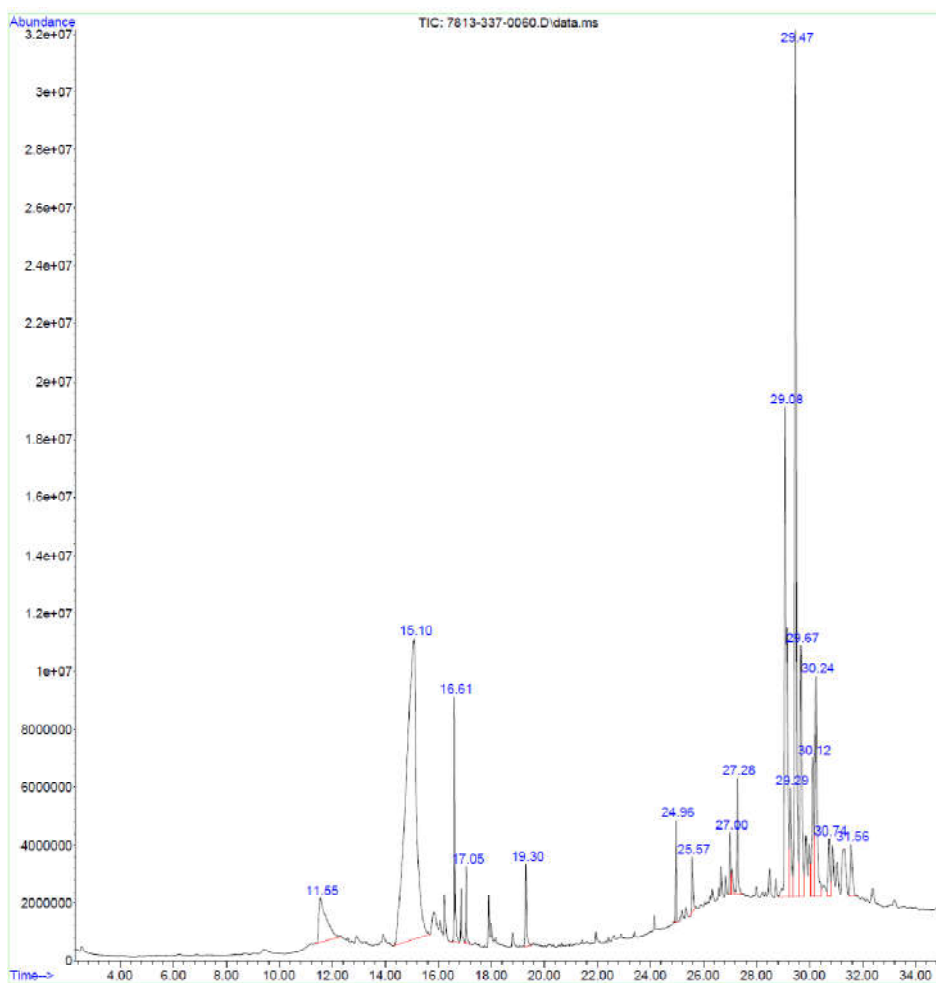


Fig. 2. Peak area of Bio-active Chemicals in Ethanol Extract of *Excoecaria agallocha* L. (GC-MS)

Table 4. Bio-active chemicals identified in *Excoecaria agallocha* L. (Ethanol extract) (GC-MS Report TLC)

Peak	R.Time(Mins)	Name of the Chemical compound	M.F.	M.W.	Peak Area	Area%	
1	16.607	Bicyclo(3,1.1)Heptane,2,6,6 Trimethyl-	(C ₁₀ H ₁₆)n	000	66904320	1.13	
2	19.308	Phytol	C ₂₀ H ₄₀ O	296	43891216	0.74	
3	24.144	Eicosane	C ₂₀ H ₃₈	278	17190394	0.29	
4	24.957	Squalene	C ₃₀ H ₅₀	410	100848316	1.70	
5	25.161	Cyclohexanone-2-2-dimethyl 5(3 methyl)	C ₈ H ₁₄ O	126	52040461	0.88	
6	25.567	Hexadecane 1-iodo-	C ₁₆ H ₃₃ I	352	219245211	3.70	
7	26.569	Oxirane,Hexadecyl	C ₁₅ H ₃₂ O	240	106771039	1.80	
8	26.657	Gamma-Tocopherol	C ₂₈ H ₄₈ O ₂	416	50760212	0.86	
9	27.005	Hentriacontane	C ₃₁ H ₆₄	436	488711916	8.25	
10	27.281	Vitamin E	C ₂₉ H ₅₀ O ₂	430	106482204	1.80	
11	28.341	16-Heptadecanal	C ₁₇ H ₃₄ O	254	151004464	2.55	
12	29.082	Tricyclo(6.2.1.0(4,11)Undecene 1,5,9,9-Tetramethyl	C ₁₅ H ₂₄	204	1182079977	19.95	
13	29.285	Naphthalene	C ₁₀ H ₁₀ O	146	184169474	3.11	
14	29.474	Tetramethyl-6,7-8,8a-Tetrahydro-5H- naphthalene-1-one	C ₁₀ H ₁₈	138	1547490440	26.12	
15	29.677	β-Amyrin	C ₃₀ H ₅₀ O	426	410800848	6.93	
16	29.866	Caparratriene	C ₁₅ H ₂₆	206	63312855	1.07	
17	30.128	Lanosterol	C ₃₀ H ₅₀ O	426	164515208	2.78	
18	30.244	α-Amyrin	C ₃₀ H ₅₀ O	426	399600429	6.74	
19	30.825	1,19-Eicosadiene	C ₂₀ H ₃₈	278	487226123	8.22	
20	31.057	Anthracene,9,10 dihydro 9,9,10 trimethyl	C ₁₇ H ₁₈	222	82299986	1.39	
					Total	6832980093	99.73

In *Wrightia tinctoria* (Roxb) R.br. Chloroform leaf extract shows inhibition on *E.coli*, *S.aureus* but not on *P.aeruginosa* (Vedhanarayanan *et al.*, 2013). It is not similar to *E.coli* with the present study. Ethanol extract of *E.agallocha* L. showed 66.7% inhibition on micro organisms used *S.aureus* was highly inhibited. *E.coli* and *B.cereus* were affected in 1000, 1500, 2000µg concentrations. *P.aeruginosa* was affected only

in 2000µg/ml. *K.pneumoniae* was affected only in the 1500 and 2000µg concentrations (Table 2 and Plate 2). Antimicrobial activity of *Eclipta alba* and *Morinda citrifolia* L. extracts of Ethanol is effective on *B.cereus*, *S.aureus*, *E.coli*, *P.aeruginosa*, *K.pneumoniae* (Mukesh *et al.*, 2010). This is not same with the results of *E.agallocha* L. Ethanol extract of *Aloe vera* showed inhibition on *K.pneumoniae*,

P. aeruginosa, *S.aureus*, but no inhibition on *E.coli* (Thiruppathi *et al.*, 2010). It is similar with *E.agallocha* L. Ethanol extract of *Citrus sinensis* shows inhibition on *E.coli*, *P.aeruginosa*, *K.pneumoniae*, *S.aureus* (Uchechi *et al.*, 2010). It is not similar with *E.agallocha* L. Ethanol extract of *Calotropis gigantea* Linn. Shows no effect on *S.aureus* and *E.coli*. It is similar with *E.agallocha* L. but active on *K.pneumoniae* (Bharathi *et al.*, 2011). The *Nelumbo nucifera* leaf extract of ethanol shows inhibition on *E.coli* and *K.pneumoniae* in 1500µg/ml (Muthu Mohammad Jamal Moideen *et al.*, 2011). It is similar with *E.agallocha* L. *Solanum nigrum* leaf extract was effective on *E.coli*, *K.pneumoniae* and *S. aureus* (Sridhar *et al.*, 2011). But this is not similar for *E.coli* and *K.pneumoniae* in *Excoecaria agallocha* L. Leaf extract of *Catharanthus roseus* (Linn) inhibits *E.coli*, *S.aureus* and *P.aeruginosa* (Chinna venkatraman *et al.*, 2012). This result is not same with *E.agallocha* L. *Casuarina equisetifolia* leaf extract inhibits very high on *S.aureus*, *E.coli*, *P.aeruginosa* and *K.pneumoniae* (Nehad and Gumgumjee *et al.*, 2012). But there was moderate effect in *E.agallocha* L. 87% of inhibition was noticed by Malarkodi and Manoharan 2013 in *Parthenium hysterophorus* extract on *B.cereus*. This is high when compared with *E.agallocha* L. *E.coli* was affected low in Ethanol extract of *Wrightia tinctoria* (Vedhanarayanan *et al.*, 2013). It is similar with *E.agallocha* Ethanol extract. In GC-MS analysis of Chloroform extract of *E.agallocha* L. 34 chemical compounds were identified (Table 3). The structural formula, molecular weight, retention time, peak area and area% were given in Figure 1. Among the 34 compounds 7 major components were high in proportion. They are Hentriacontane 5.23%, Tricyclo, undec-tetramethyl 16.38%, Tetramethyl, trihydro-naphthalene 20.72%, β - amyryn 7.52%, 9,19-cyclolanost :24-en-3-ol 4.47%, α - amyryn 6.92%, Heptadecanol 8.24%. All the constituents were characterized and identified by comparison of the mass spectra of the constituents of the known components stored in NIST library. The presence of various bio-active compounds in *E.agallocha* justifies the use of leaves for various ailments traditionally. Hentriacontane is in bees wax also (8.9%). In Ethanol extract of *E.agallocha* L. seventeen compounds were identified (Table 4 and Fig.2). From these Myoinosital 4-c-methyl 37.09%, Tricyclo undec(isocaryophyllene) 13.21%, Trimethyl 5,6-dimethylene-deca hydro naphthalene 17.25%, β - amyryn 5.78%, α - amyryn 5.78% were major constituents.

Conclusion

This investigation shows that *E.agallocha* L. has lot of antibacterial activity against many bacteria responsible for most of the microbial diseases. GC-MS analysis of Chloroform extract of *E.agallocha* shows 34 bio-active compounds. From that seven compounds are high in proportion. They are very useful for various ailments and diseases. Ethanol extract shows 17 compounds in that 5 compounds, are high in proportion. They have broad spectrum of use. It is used in perfume, food stuffs and beverages. Cyclolanost is an important component in Dandelion coffee. It is very good tonic for liver. Jaundice affect the liver severely. For the jaundice patients, the tonic is very useful. α -amyryn and β - amyryn are used as laxative and used in cancer treatment. Some compounds are used in cleaning agents and used as dying solubilizing agent in textiles. Further studies are needed for the isolation and identification of bio-active compounds using some other

solvents and micro organisms gives better understanding of bio- active compounds.

REFERENCES

- Agwa, A. *et al.*, 2000. "Isolation And Characterization Of Two Streptomyces Species Produced Non Polyenic Antifungal Agents ." *Journal Of Union Arab Biology* 7 : 62: 82.
- Asirvadam doss and Rangaswamy dhanabalan, 2008. "Preliminary Phytochemical Screening and Antibacterial studies of Leaf extract of *Solanum trilobatum* Linn." *Ethanobotanical leaflets* 12:638-42.
- Bandaranayake, W.M. 2002. "Bioactivities, bioactive compounds and chemical constituents of mangrove plant wet land Ecology and management."10 (32): 421:452.
- Bharathi, P. *et al.*, 2011. " Antibacterial activity of leaf extracts of *Calotropis gigantea* Linn. against certain gram negative and gram positive bacteria" *International Journal of Chemical Sciences*, 9(2) : 919-923.
- Chinna venkatraman, *et al.* 2012. "Invitro Antibacterial Activity And Phytochemical Analysis Of *Catharanthus roseus* Linn." *Asian Pacific Journal of Tropical Bio Medicines*, 5155-5158.
- Gamble J.S. 1954. "Flora of Presidency of Madras". Volume . II : 940
- Jayanta Kumar Patra *et al.* 2009. "Phytochemical screening and antimicrobial assessment of leaf extract of *E. agallocha* L. " *Advances in Natural and Applied Sciences*. 3(2): 241-246).
- Malarkodi, E. and Manoharan, A. 2013. "Study on antibacterial activity of *Parthenium hysterophorus* L" *Journal of Chemical and Pharmaceutical Research*, 5(1) : 134-136.
- Mukesh Chandra Sharma and Smita Sharma 2010 " Phytochemical screening of Methanol extract and antibacterial activity of *Eclipta alba* and *Morinda citrifolia* L." *Middle - east Journal of Scientific Reasearch* 6(5): 445-449.
- Muthu Mohammad *et al.* 2011. " Antimicrobial activity and Phytochemical analysis of *Nelumbo nucifere* leaves" *Journal of Global Trends in Pharmaceutical Sciences* as volume 2 : 404-410.
- Nehad, M. and Gumgumjee, *et al.* 2012. "Antibacterial efficacy of *Casuarina equisetifolia* extracts against some pathogenic micro organisms ." *Journal of Medicinal Plant Research*, Vol. 6(47) : 5819 -5825.
- Perez, C. *et al.* 1990. "An antibiotic assay by the agar well diffusion method". *Acta.Bio.Med.Exp.*, 15:113-115.
- Peter, K.L.N *et al.* 1999. "Guide to the Mangrove of Singapore I: The Ecosystem and Plant Diversity, "Singapore Science Centre : 111-112.
- Sridhar, T.M. *et al.* 2011. "Invitro Antibacterial activity and Phytochemical analysis of *Solanum nigrum* Linn. an important antiulcer medicinal plant" *Journal of Experimental Sciences*, 2(8). 24-29.
- Thiruppathi, S. *et.al.* 2010. "Antibacterial activity of *Aloe vera* (L.) Burm.F against Pathogenic micro organisms" *Journal of Biological Sciences Research*, vol 1(4) 251-258.
- Uchechi, N. 2010. "The antibacterial activity of crude leaf extract of *Citrus sinensis* (sweet orange)" *Inter. Journal of Pharma and Bio-Sciences*, Vol 1./issue 4 /Oct-Dec-2010.
- Vedhanarayanan, P. *et al.* 2013. "Antimicrobial activity and Phytochemical screening of *Wrightia tinctoria* (Roxb) R.Br," *Journal of Pharmacognosy and Phytochemistry.*, 2(4). 123-125.