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REVIEW ARTICLE

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NANOTECHNOLOGY IN VASCULAR MEDICINE A REVIEW

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ABSTRACT

Although the application of materials produced in nanotechnology with a size of 1-100 nm in at least one of their dimensions at the molecular level opens new perspectives in the care of patients, the question of cost-effectiveness and the safety of nanotechnology still remain open. These have proven to be useful not only as prosthetic materials, but also for surface preparation of implants and prostheses, effective drug delivery systems for antibiotics, and chemotherapeutics, and drug eluting systems to combat implant-related infections, e.g.. The application of nanotechnology in vascular medicine firstly extends to both drug-eluting therapies for obstructive vascular diseases and prosthetic materials used for surgical revascularization. The novel nanomaterials can deliver the thrombolytic drugs directly to the lesion.

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INTRODUCTION

Although the application of materials produced in nanotechnology with a size of 1-100 nm in at least one of their dimensions at the molecular level opens new perspectives in the care of patients, the question of cost-effectiveness and the safety of nanotechnology still remain open (1). These have proven to be useful not only as prosthetic materials, but also for surface preparation of implants and prostheses, effective drug delivery systems for antibiotics, and chemotherapeutics, and drug eluting systems to combat implant-related infections, etc. (2-4). The application of nanotechnology in vascular medicine firstly extends to both drug-eluting therapies for obstructive vascular diseases and prosthetic materials used for surgical revascularization. The novel nanomaterials can deliver the thrombolytic drugs directly to the lesion (5).

Aim of the review: In the near future, new developments in nanotechnology will make it possible to therapeutically address the underlying mechanisms of atherosclerosis directly at their point of origin and to validly measure the success of therapy using the same method. In doing so, the complications to be feared from systemic therapy can be avoided and the therapeutic dose even reduced. The present work deals with the various facets of the new developments in nanotechnology in the diagnosis and therapy of vascular diseases, especially atherosclerosis.

METHODS

This systematic review was performed using available databases: PubMed, Medline, Cochrane Library, Embase, and Clinical Trials.gov. Unpublished data remained unconsidered. The keywords Key words entered were nanotechnology, vascular disease, molecular biology basis of atherosclerotic plaques, nanostents, nanoprostheses. Only studies that showed a clear result were considered. Pilot studies were not considered.

DISCUSSION

Our current drug therapies are characterized by poor target specificity and too low delivery efficiency (6-8). By altering their shape, size, and surface chemistry, the site-specific delivery of nanoparticles can be specifically programmed (9,10). As a first particle, the use of Doxil, a liposomally encapsulated doxorubicin formulation for the treatment of Kaposi's sarcoma significantly reduced doxorubicin-induced cardiotoxicity and heart failure typical of this treatment by cancer-specific toxicity (11,12). Other nanoparticle drug delivery systems developed on this basis are used in infections, chronic kidney disease, e.g. (13). In vascular medicine, nanoformulations of fenofibrate are used in patients with hypertriglyceridemia. This has completely abolished significant problems associated with the

ingestion of fenofibrates, such as solubility and absorption (14-16). Promising are the new developments with additional nanoparticles for treatment with Annexin A1 mimetic peptide and IL-10 bound to Type IV collagen-targeted copolymers of PLGA-PEG, superoxide dismutase mimetic agent and hydrogen peroxide-eliminating compound, bound to cyclodextrin-based polysaccharide, prednisolone bound to MRI-detectable liposomes, VCAM1, ICAM1 and 2, E- and P-selectin siRNA bound to PEI polymer as some examples (17-29). Appropriate nanoparticles are also being tested for supraselective thrombolytic therapy with streptokinase, urokinase, or thrombin inhibitors (30-35).

The inflammation typical of atherosclerosis is maintained by a lack of programmed cellular clearance of apoptotic cells (36-38). Their suppression by canakinumab carries the risk of decreased innate immunity and increased mortality (39-41). Nanoparticles acting only on atherosclerotic plaques may minimize this risk (41). The principle of action of this procedure can be attributed to the specific function of type IV collagen, the release of which plays a central role in vascular injury and inflammation (42). Binding of annexin A1-derived peptide to a type IV collagen targeting peptide resulted in a 70% increase in nanoparticle selectivity for atherosclerotic lesions (43). A similar effect was achieved by binding anti-inflammatory cytokine IL (interleukin)-10 to type IV collagen-targeted copolymers of PLGA-PEG (18). In addition, nanoparticles can be used theranostically (therapeutic-diagnostic). In animal studies, binding of prednisolone to an MRI-detectable liposome prolonged its half-life, without systemic toxicity, and demonstrated a sustained decrease in plaque inflammation on 18F-FDG positron emission tomography/computed tomography (44-46).

In another animal study, binding of small interfering RNA (siRNA) directed against multiple adhesion molecules to a polymer-based nanoparticle significantly reduced tissue damage and necrotic core formation after coronary ligation following an ischemic insult (47,48). Neointimal neovascularization significantly correlates with increased plaque instability and subsequent symptoms (49-51). This process is triggered by VEGF (vascular endothelial growth factor) and platelet-derived growth factor, among others. Anti-VEGF therapy, mainly by binding VCAM1, ICAM1 and 2, E- and P-selectin siRNA to a PEI polymer, reduced plaque development in apoE^{-/-} mice. At the same time, the MRI-detectable nanoparticles allowed the T1-weighted MRI signal to be measured in the aorta as a parameter of inflammation and thus could be evaluated diagnostically (47,52-57). A central role in the pathogenesis of atherosclerosis beyond lesion initiation is played by macrophages, especially in misdirection of apoptotic cells in terms of efferocytic activity (58,59). Several nanotherapies target monocyte recruitment and infiltration in plaque, proliferation of macrophages with polarization to a less inflammatory M2 phenotype, and cholesterol metabolism (14-16,22,25,60). Nanoparticles were also shown to inhibit the uptake of oxidized LDL by macrophage SRs (scavenger receptors), resulting in a reduction of lipid load and thereby decreased reduced plaque occlusion in the aorta of ApoE^{-/-} mice (24).

Binding of TRAF6 inhibitors into recombinant high-density lipoprotein (HDL) nanoparticles (TRAF6i-HDL) blocked cluster of differentiation 40 (CD40)-induced TRAF6 (tumor necrosis receptor-associated factor 6) in monocytes and macrophages, resulting in stable plaque phenotypes and no adverse immune responses (18,26,61). The combination of nanotherapy with phototherapy gave rise to controversial discussions. Phototherapy with near-infrared fluorophore of inflammatory monocytes and macrophages that have previously ingested iron oxide nanoparticles not infrequently results in ablation of macrophage-rich plaques in animal studies (62,63). This may increase the risk of plaque rupture (64). Precisely to avoid this highly dangerous side effect, cell-specific single-walled carbon nanotubes (SWNTs) that are highly selectively taken up by inflammatory Ly-6Chi monocytes are increasingly being developed (65). In this context, their natural photoacoustic contrast and a near-infrared fluorescence signal open additional diagnostic possibilities of SWNTs (66). Anti-stenotic agents used for targeted inhibition of

restenosis after peripheral revascularization can act directly on the treated vascular bed by binding to nanoparticles (67,68). For example, supraselective endovascular delivery of albumin-bound rapamycin nanoparticles reduced luminal stenosis after balloon angioplasty of the femoral artery in a porcine model (69). Similar results were obtained in another study using $\alpha v\beta 3$ -targeted paramagnetic nanoparticles for the delivery of rapamycin (70-72). In the new generations of drug-eluting stents, paclitaxel bound to nanoparticles is attached to surfaces of stents (73). This has already yielded promising results in animal studies (74-76). Thus, even much higher doses of paclitaxel could be tolerated (77-80). Similar results were obtained in animal studies as well as in clinical trials with liposomal formulation of the bisphosphonate alendronate. Although a significant difference in restenosis rate was found between the treatment and placebo groups, the rate of in-stent late loss was significantly lower in patients with an elevated monocyte count at baseline (81). One of the major challenges in interventional therapy is in-stent restenosis. The restenosis rate of 40% in the treatment of coronary artery disease with a drug-eluting stent and in the treatment of femoral artery stenosis after only 24 months was not different from that of the placebo group of 44% (82). In addition, the paclitaxel- and sirolimus-eluting stents exhibit significantly higher rates of thrombosis via slowed endothelialization, which can be lethal, especially with poly-n-butyl methacrylate- and polyethylene-vinyl acetate copolymer-eluting prostheses (83-85). Microfabricated drug-release reservoirs used in two new stents (the Janus CarboStent, Sorin Biomedica Cardio S.p.A., Via Crescentino, Italy, and the Conor Stent, Conor Medsystems, Inc, Menlo Park, California) are promising (86).

In this context, neointima hyperplasia is affected more by a prolonged release phase than by the dose itself (87,88). Future directions include stents fabricated by microelectroerosion machining (μ EDM) (89,90). A different manufacturing technique using sharp silicon microneedles with a height of 80 to 140 μ m allow local accumulation of therapeutic agents by penetrating dense atherosclerotic lesions (91). Technical difficulties hamper the implementation of this development (92,93). As a new generation of medicinal agents as nanoscale texture hydroxyapatite and titanium oxide in development (94-98). Electrospun poly ϵ -caprolactone nanofiber scaffold (PCL), which is hydrophobic due to surface modification with gelatin and architecturally mimics ECM, represent new generation of vascular prostheses (99-105). The poor long-term results caused by lack of geometric alignment of previously used prostheses such as PTFE and Dacron could be compensated by the development of electrospun scaffolds (ES) (106-110). Endothelial injury induced during angioplasty represents one of the main initiators of intimal hyperplasia, which could be reduced in a dose-dependent manner by doxorubicin-loaded nanoparticles in a rat model (111,112). Polyhedral oligomeric silsesquioxane (POSS) and polyhedral oligomeric silsesquioxane poly (carbonate-urea)-urethane (POSS-PCU) used to coat new generation prostheses are characterized by their antithrombotic properties as well as stimulation of endothelialization (113-115). Also in valve replacement surgery, polymeric heart valves (PHVs) represent an alternative to existing prostheses whose use in routine clinical practice will certainly take years (116-119). Based on a functionalized graphene oxide (FGO) nanomaterials in a poly(carbonate urea) urethane (PCU) backbone, the nanotechnology-based prosthesis, Hastalex has shown good results in vivo trials (120).

Conclusions

The multiple developments in nanotechnology not only enable a new therapeutic approach to specifically modify atherosclerotic plaque at the molecular level, but at the same time allow verification of therapeutic efficacy. As a result, non-lower single therapeutic doses of drugs can be administered. The side effects occurring in the process can also be reduced to a minimum. The new generation of bioprostheses based on nanotechnology will revolutionize the long-term results of vascular interventions.

List of abbreviations

VCAM1: Vascular Cell Adhesion Protein 1
ICAM1: Intercellular Adhesion Molecule
PEI polymer: Polyethylenimin polymer
PLGA-PEG: poly-Lactide-Co-Glycolide A-PpolyEthylene Glycol
VEGF: Vascular Endothelial Growth Factor
MRI: Magnet Resonance Imaging
TRAF6: tumor necrosis receptor-associated factor 6
SWNTs: single-walled carbon nanotubes
μEDM: microelectroerosion machining
PCL: poly ε-caprolactone nanofiber scaffold
ECM: extracellular matrix
PTFE: Polytetrafluorethylen
ES: electrospun scaffolds
POSS: Polyhedral oligomeric silsesquioxane
POSS-PCU: polyhedral oligomeric silsesquioxane poly(carbonate-urea)-urethane
PHVs: polymeric heart valves
FGO: functionalized graphene oxide

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REFERENCES

- Johnson R, Ding Y, Nagiah N, Monnet E, Tan W. Coaxially-structured fibres with tailored material properties for vascular graft implant. *Mater Sci Eng C Mater Biol Appl.* 2019; 97: 1–11. doi: 10.1016/j.msec.2018.11.036.
- Adiguzel E, Ahmad PJ, Franco C, Bendeck MP. Collagens in the progression and complications of atherosclerosis. *Vasc Med.* 2009; 14:73–89. doi: 10.1177/1358863X08094801.
- Al-Enizi AM, Zagho MM, Elzatahry AA. Polymer-Based Electrospun Nanofibers for Biomedical Applications. *Nanomaterials.* 2018; 8: 259. doi: 10.3390/nano8040259.
- Aoki J, Ong AT, Granillo GAR et al. “Full metal jacket” (stented length > or =64 mm) using drug-eluting stents for de novo coronary artery lesions. *Am Heart J.* 2005; 150: 994-999. doi: 10.1016/j.ahj.2005.01.050.
- Areva S, Paldan H, Peltola T, Narhi T, Jokinen M, Linden M. Use of sol-gel-derived titania coating for direct soft tissue attachment. *J Biomed Mater Res A.* 2004; 70:169-178. doi: 10.1002/jbm.a.20120.
- Banai S, Finkelstein A, Almagor Y et al. Targeted anti-inflammatory systemic therapy for restenosis: the Biorest Liposomal Alendronate With Stenting Study (BLAST)-a double blind, randomized clinical trial. *Am Heart J.* 2013; 165:234.e1–40.e1. doi: 10.1016/j.ahj.2012.10.023.
- Barenholz Y. Doxil®—the first FDA-approved nano-drug: lessons learned. *J Control Release.* 2012; 160:117–134. doi: 10.1016/j.jconrel.2012.03.020.
- Bate TSR, Forbes SJ, Callanan, A. J. Controlling Electrospun Polymer Morphology for Tissue Engineering Demonstrated Using hepG2 Cell Line. *Vis Exp.* 2020; 159: e61043. doi: 10.3791/61043.
- Baylis RA, Gomez D, Mallat Z, Pasterkamp G, Owens GK. The CANTOS trial: one important step for clinical cardiology but a giant leap for vascular biology. *Arterioscler Thromb Vasc Biol.* 2017; 37:e174–e177. doi: 10.1161/ATVBAHA.117.310097.
- Bishop JA, Palanca AA, Bellino MJ, Lowenberg DW. Assessment of compromised fracture healing. *J Am Acad Orthop Surg.* 2012;20(5):273-82. doi: 10.5435/JAAOS-20-05-273.
- Bucci R, Vaghi F, Erba E, Romanelli A, Gelmi ML, Clerici F. Peptide grafting strategies before and after electrospinning of nanofibers. *Acta Biomater.* 2021; 1;122:82-100. doi: 10.1016/j.actbio.2020.11.051.
- Caves JM, Chaikof EL. The evolving impact of microfabrication and nanotechnology on stent design. Author links open overlay panel. *J Vasc Surg.* 2006; 44(6): 1363-1368. doi: 10.1016/j.jvs.2006.08.046.
- Chan JM, Rhee JW, Drum CL et al. In vivo prevention of arterial restenosis with paclitaxel-encapsulated targeted lipid-polymeric nanoparticles. *Proc Natl Acad Sci USA.* 2011; 108:19347–19352. doi: 10.1073/pnas.1115945108.
- Chorny M, Fishbein I, Yellen BB et al. Targeting stents with local delivery of paclitaxel-loaded magnetic nanoparticles using uniform fields. *Proc Natl Acad Sci USA.* 2010;107:8346–8351. doi: 10.1073/pnas.0909506107.
- Chorny M, Fishbein I, Yellen BB et al. Targeting stents with local delivery of paclitaxel-loaded magnetic nanoparticles using uniform fields. *Proc Natl Acad Sci USA.* 2010; 107:8346–8351. doi: 10.1073/pnas.0909506107.
- Cui Wenguo, Wang Aijun, Zhao Chao, Zhu Wuqiang. Nanotechnology in Cardiovascular Regenerative Medicine. *Front Bioeng Biotechnol.* 2020; 8: 608844. doi: 10.3389/fbioe.2020.608844.
- Cyrus T, Zhang H, Allen JS et al. Intramural delivery of rapamycin with alphavbeta3-targeted paramagnetic nanoparticles inhibits stenosis after balloon injury. *Arterioscler Thromb Vasc Biol.* 2008; 28:820–826. doi: 10.1161/ATVBAHA.107.156281.
- De la Zerd A, Zavaleta C, Keren S et al. Carbon nanotubes as photoacoustic molecular imaging agents in living mice. *Nat Nanotechnol.* 2008; 3:557–562. doi: 10.1038/nnano.2008.231.
- Duda SH, Bosiers M, Lammer J et al. Sirolimus-eluting versus bare nitinol stent for obstructive superficial femoral artery disease: the SIROCCO II trial. *J Vasc Interv Radiol.* 2005; 16: 331-338. doi: 10.1097/01.RVI.0000151260.74519.CA.
- Duivenvoorden R, Tang J, Cormode DP et al. A statin-loaded reconstituted high-density lipoprotein nanoparticle inhibits atherosclerotic plaque inflammation. *Nat Commun.* 2014; 5:3065. doi: 10.1038/ncomms4065
- Dulak J, Józkwicz A. Anti-angiogenic and anti-inflammatory effects of statins: relevance to anti-cancer therapy. *Curr Cancer Drug Targets.* 2005; 5:579–594.
- Duncan R. The dawning era of polymer therapeutics. *Nat Rev Drug Discov.* 2003; 2:347–360. doi: 10.1038/nrd1088.
- Dunmore BJ, McCarthy MJ, Naylor AR, Brindle NP. Carotid plaque instability and ischemic symptoms are linked to immaturity of microvessels within plaques. *J Vasc Surg.* 2007; 45:155–159. doi: 10.1016/j.jvs.2006.08.072.
- Endo M, Koyama S, Matsuda Y, Hayashi T and Kim YA. Thrombogenicity and blood coagulation of a microcatheter prepared from carbon nanotube-nylon-based composite. *Nano Lett.* 2005; 5(1):101-5. doi: 10.1021/nl0482635.
- Escudier B, Eisen T, Stadler WM et al. Sorafenib for treatment of renal cell carcinoma: final efficacy and safety results of the phase III treatment approaches in renal cancer global evaluation trial. *J Clin Oncol.* 2009; 27:3312–3318. doi: 10.1200/JCO.2008.19.5511.
- Finkelstein A, McClean D, Kar S et al. Local drug delivery via a coronary stent with programmable release pharmacokinetics. *Circulation.* 2003; 107: 777-784. doi: 10.1161/01.cir.0000050367.65079.71.
- Flores AM, Ye J, Jarr KU, Hosseini-Nassab N, Smith BR, Leeper NJ. Nanoparticle Therapy for Vascular Diseases. *Arterioscler Thromb Vasc Biol.* 2019; 39(4): 635–646. doi: 10.1161/ATVBAHA.118.311569.
- Fordyce CB, Roe MT, Ahmad T, et al. Cardiovascular drug development: is it dead or just hibernating? *J Am Coll Cardiol.* 2015;65:1567–1582. doi: 10.1016/j.jacc.2015.03.016.
- Frank-Kamenetsky M, Grefhorst A, Anderson NN et al. Therapeutic RNAi targeting PCSK9 acutely lowers plasma cholesterol in rodents and LDL cholesterol in nonhuman primates. *Proc Natl*

- Acad Sci USA. 2008;105:11915–11920 doi: 10.1073/pnas.0805434105.
- Fredman G, Kamaly N, Spolitu S et al. Targeted nanoparticles containing the proresolving peptide Ac2-26 protect against advanced atherosclerosis in hypercholesterolemic mice. *Sci Transl Med.* 2015; 7:275ra20. doi: 10.1126/scitranslmed.aaa1065.
- Fredman G, Kamaly N, Spolitu S et al. Targeted nanoparticles containing the proresolving peptide Ac2-26 protect against advanced atherosclerosis in hypercholesterolemic mice. *Sci Transl Med.* 2015; 7:275ra20. doi: 10.1126/scitranslmed.aaa1065.
- Gabizon A, Shmeeda H, Barenholz Y. Pharmacokinetics of pegylated liposomal Doxorubicin: review of animal and human studies. *Clin Pharmacokinet.* 2003;42:419-436. doi: 10.2165/00003088-200342050-00002.
- Gao W, Sun Y, Cai M et al. Copper sulfide nanoparticles as a photothermal switch for TRPV1 signaling to attenuate atherosclerosis. *Nat Commun.* 2018; 9:231. doi: 10.1038/s41467-017-02657-z.
- Gasper WJ, Jimenez CA, Walker J, Conte MS, Seward K, Owens CD. Adventitial nab-rapamycin injection reduces porcine femoral artery luminal stenosis induced by balloon angioplasty via inhibition of medial proliferation and adventitial inflammation. *Circ Cardiovasc Interv.* 2013; 6:701–709. doi: 10.1161/CIRCINTERVENTIONS.113.000195.
- Gavaskar A, Rojas D, Videla F. Nanotechnology: the scope and potential applications in orthopedic surgery. *Eur J Orthop Surg Traumatol.* 2018;28:1257-1260. doi.org/10.1007/s00590-018-2193-z.
- Ghanbari H, Viatge H, Kidane AG, Burriesci G, Tavakoli M, Seifalian AM. Polymeric heart valves: new materials, emerging hopes. *Trends Biotechnol.* 2009;27(6): 359-367. doi: 10.1016/j.tibtech.2009.03.002.
- Gössl M, Versari D, Hildebrandt HA et al. Segmental heterogeneity of vasa vasorum neovascularization in human coronary atherosclerosis. *JACC Cardiovasc Imaging.* 2010; 3:32–40. doi: 10.1016/j.jcmg.2009.10.009.
- Greenberg JI, Shields DJ, Barillas SG et al. A role for VEGF as a negative regulator of pericyte function and vessel maturation. *Nature.* 2008; 456:809–813. doi: 10.1038/nature07424.
- Guo L, Harari E, Virmani R, Finn AV. Linking hemorrhage, angiogenesis, macrophages, and iron metabolism in atherosclerotic vascular diseases. *Arterioscler Thromb Vasc Biol.* 2017; 37:e33–e39. doi: 10.1161/ATVBAHA.117.309045.
- Hanc M, Fokter SK, Vogrin M, Vogrin M, Molicnik A, Recnik G. Porous tantalum in spinal surgery: an overview. *Eur J Orthop Surg Traumatol.* 2016; 26(1):1-7. doi: 10.1007/s00590-015-1654-x.
- He Z, Rault F, Lewandowski M, Mohsenzadeh E, Salaün F. Electrospun PVDF Nanofibers for Piezoelectric Applications: A Review of the Influence of Electrospinning Parameters on the beta Phase and Crystallinity Enhancement. *Polymers.* 2021; 13(2): 174. doi.org/10.3390/polym13020174.
- Henry S, McAllister DV, Allen MG, Prausnitz MR. Microfabricated microneedles: a novel approach to transdermal drug delivery. *J Pharm Sci.* 1998; 87: 922-925. doi: 10.1021/js980042+.
- Huang SL, Kee PH, Kim H et al. Nitric oxide-loaded echogenic liposomes for nitric oxide delivery and inhibition of intimal hyperplasia. *J Am Coll Cardiol.* 2009; 54:652–659. doi: 10.1016/j.jacc.2009.04.039.
- Huang SL, Kee PH, Kim H et al. Nitric oxide-loaded echogenic liposomes for nitric oxide delivery and inhibition of intimal hyperplasia. *J Am Coll Cardiol.* 2009; 54:652–659. doi: 10.1016/j.jacc.2009.04.039.
- Hurwitz H, Fehrenbacher L, Novotny W et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N Engl J Med.* 2004; 350:2335–2342. doi: 10.1056/NEJMoa032691.
- Iakovou I, Schmidt T, Bonizzoni E et al. Incidence, predictors, and outcome of thrombosis after successful implantation of drug-eluting stents. *JAMA.* 2005(293): 2126-2130. doi: 10.1001/jama.293.17.2126.
- Jaganathan, S K, Mani MP. Enriched mechanical, thermal, and blood compatibility of single stage electrospun polyurethane nickel oxide nanocomposite for cardiac tissue engineering. *Polym Compos.* 2019; 40(6): 2381-2390. doi:10.1002/pc.25098.
- Kamaly N, Fredman G, Fojas JJ et al. Targeted interleukin-10 nanotherapeutics developed with a microfluidic chip enhance resolution of inflammation in advanced atherosclerosis. *ACS Nano.* 2016; 10:5280–5292. doi: 10.1021/acsnano.6b01114.
- Kamba T, McDonald DM. Mechanisms of adverse effects of anti-VEGF therapy for cancer. *Br J Cancer.* 2007; 96:1788–1795. doi: 10.1038/sj.bjc.6603813.
- Kasikara C, Doran AC, Cai B, Tabas I. The role of non-resolving inflammation in atherosclerosis. *J Clin Invest.* 2018; 128:2713–2723. doi: 10.1172/JCI97950.
- Katsuki S, Matoba T, Nakashiro S, et al. Nanoparticle-mediated delivery of pitavastatin inhibits atherosclerotic plaque destabilization/rupture in mice by regulating the recruitment of inflammatory monocytes. *Circulation.* 2014;129:896–906. doi: 10.1161/CIRCULATIONAHA.113.002870.
- Kienapfel H, Sprey C, Wilke A, Griss P. Implant fixation by bone ingrowth. *J Arthroplasty.* 1999;14(3):355-68. doi: 10.1016/s0883-5403(99)90063-3.
- Kojima Y, Downing K, Kundu R et al. Cyclin-dependent kinase inhibitor 2B regulates efferocytosis and atherosclerosis. *J Clin Invest.* 2014; 124:1083–1097. doi: 10.1172/JCI170391.
- Kojima Y, Weissman IL, Leeper NJ. The role of efferocytosis in atherosclerosis. *Circulation.* 2017; 135:476–489. doi: 10.1161/CIRCULATIONAHA.116.025684.
- Kolodgie FD, John M, Khurana C et al. Sustained reduction of in-stent neointimal growth with the use of a novel systemic nanoparticle paclitaxel. *Circulation.* 2002; 106:1195–1198.
- Kolodgie FD, John M, Khurana C et al. Sustained reduction of in-stent neointimal growth with the use of a novel systemic nanoparticle paclitaxel. *Circulation.* 2002; 106:1195–11. doi: 10.1161/01.cir.0000032141.31476.15.
- Kolodgie FD, John M, Khurana C et al. Sustained reduction of in-stent neointimal growth with the use of a novel systemic nanoparticle paclitaxel. *Circulation.* 2002; 106: 1195-1198. doi: 10.1161/01.cir.0000032141.31476.15.
- Kosuge H, Sherlock SP, Kitagawa T et al. Near infrared imaging and photothermal ablation of vascular inflammation using single-walled carbon nanotubes. *J Am Heart Assoc.* 2012; 1:e002568. doi: 10.1161/JAHA.112.002568.
- Kotani J, Awata M, Nanto S et al. Incomplete neointimal coverage of sirolimus-eluting stents: angioscopic findings. *J Am Coll Cardiol.* 2006; (47): 2108-2111. doi: 10.1016/j.jacc.2005.11.092.
- Lameijer M, Binderup T, van Leent M et al. Efficacy and safety assessment of a TRAF6-targeted nanoimmunotherapy in atherosclerotic mice and non-human primates. *Nat Biomed Eng.* 2018; 2:279–292.
- Leuschner F, Dutta P, Gorbato R et al. Therapeutic siRNA silencing in inflammatory monocytes in mice. *Nat Biotechnol.* 2011; 29:1005–1010. doi: 10.1038/nbt.1989.
- Lewis DR, Petersen LK, York AW et al. Sugar-based amphiphilic nanoparticles arrest atherosclerosis in vivo. *Proc Natl Acad Sci USA.* 2015; 112:2693–2698. doi: 10.1073/pnas.1424594112.
- Li JM, Newburger PE, Gounis MJ et al. Local arterial nanoparticle delivery of siRNA for NOX2 knockdown to prevent restenosis in an atherosclerotic rat model. *Gene Ther.* 2010; 17:1279–1287. doi: 10.1038/gt.2010.69.
- Li JM, Newburger PE, Gounis MJ, Dargon P, Zhang X, Messina LM. Local arterial nanoparticle delivery of siRNA for NOX2 knockdown to prevent restenosis in an atherosclerotic rat model. *Gene Ther.* 2010; 17:1279–1287. doi: 10.1038/gt.2010.69.
- Li, Z.; Li, X.; Xu, T.; Zhang, L. Acellular Small-Diameter Tissue-Engineered Vascular Grafts. *Appl Sci.* 2019; 9(14): 2864. doi.org/10.3390/app9142864.

- Libby P. Inflammation in atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2012; 32:2045–2051. doi: 10.1161/ATVBAHA.108.179705.
- Liu DM, Troczynski T, Tseng WJ. Water-based sol-gel synthesis of hydroxyapatite: process development. *Biomaterials.* 2001; 22: 1721-1730. doi: 10.1016/s0142-9612(00)00332-x.
- Liu DM, Yang Q, Troczynski T. Sol-gel hydroxyapatite coatings on stainless steel substrates. *Biomaterials.* 2002;23: 691-698. doi: 10.1016/s0142-9612(01)00157-0.
- Lobatto ME, Calcagno C, Otten MJ et al. Pharmaceutical development and preclinical evaluation of a GMP-grade anti-inflammatory nanotherapy. *Nanomedicine.* 2015; 11:1133–1140. doi: 10.1016/j.nano.2015.02.020.
- Mallat Z. Macrophages. *Arterioscler Thromb Vasc Biol.* 2017; 37:e92–e98. doi: 10.1161/ATVBAHA.117.309730.
- Mani MP, Jaganathan SK., Prabhakaran P, Nageswaran G, Krishnasamy NP. Fabrication and characterization of polyurethane patch loaded with palmarosa and cobalt nitrate for cardiac tissue engineering. *Int J Poly Ana Ch.* 2019; 24, 399–411. doi:10.1080/1023666x.2019.1598665.
- Margolis J, McDonald J, Heuser R et al. Systemic nanoparticle paclitaxel (nab-paclitaxel) for in-stent restenosis I (SNAPIST-I): a first-in-human safety and dose-finding study. *Clin Cardiol.* 2007; 30:165–170. doi: 10.1002/clc.20066.
- Mariappan N. Recent trends in Nanotechnology applications in surgical specialties and orthopedic surgery. *Biomed Pharmacol J.* 2019;12(3). doi:org/10.13005/bpi/1739.https://dx.doi.org/10.13005/bpi/1739.
- Masuda S, Nakano K, Funakoshi K et al. Imatinib mesylate-incorporated nanoparticle-eluting stent attenuates in-stent neointimal formation in porcine coronary arteries. *J Atheroscler Thromb.* 2011; 18:1043–1053. doi: 10.5551/jat.8730.
- Masuda S, Nakano K, Funakoshi K et al. Imatinib mesylate-incorporated nanoparticle-eluting stent attenuates in-stent neointimal formation in porcine coronary arteries. *J Atheroscler Thromb.* 2011; 18:1043–1053. doi: 10.5551/jat.8730.
- McAllister DV, Wang PM, Davis SP et al. Microfabricated needles for transdermal delivery of macromolecules and nanoparticles: fabrication methods and transport studies. *Proc Natl Acad Sci U S A.* 2003; 100: 13755-13760. doi: 10.1073/pnas.2331316100.
- McCarthy JR, Korngold E, Weissleder R, Jaffer FA. A light-activated theranostic nanoagent for targeted macrophage ablation in inflammatory atherosclerosis. *Small.* 2010; 6:2041–2049. doi: 10.1002/sml.201000596.
- Miller DC, Haberstroh KM, Webster TJ. Mechanism(s) of increased vascular cell adhesion on nanostructured poly(lactic-co-glycolic acid) films. *J Biomed Mater Res A.* 2005;73: 476-484. doi: 10.1002/jbma.a.30318.
- Miller DC, Thapa A, Haberstroh KM, Webster TJ. Endothelial and vascular smooth muscle cell function on poly(lactic-co-glycolic acid) with nano-structured surface features. *Biomaterials.* 2004;25: 53-61. doi: 10.1016/s0142-9612(03)00471-x.
- Moore KJ, Tabas I. Macrophages in the pathogenesis of atherosclerosis. *Cell.* 2011; 145:341–355. doi: 10.1016/j.cell.2011.04.005.
- Murali M, Yeo SH. Rapid biocompatible micro device fabrication by micro electro-discharge machining. *Biomed Microdevices.* 2004; 6: 41-45. doi: 10.1023/b:bmmd.0000013364.71148.51.
- Nahrendorf M, Zhang H, Hembrador S et al. Nanoparticle PET-CT imaging of macrophages in inflammatory atherosclerosis. *Circulation.* 2008; 117:379–387. doi: 10.1161/CIRCULATIONAHA.107.741181.
- Nakano K, Egashira K, Masuda S et al. Formulation of nanoparticle-eluting stents by a cationic electrodeposition coating technology: efficient nano-drug delivery via bioabsorbable polymeric nanoparticle-eluting stents in porcine coronary arteries. *JACC Cardiovasc Interv.* 2009; 2:277–283. doi: 10.1016/j.jcin.2008.08.023.
- Nakashiro S, Matoba T, Umezumi R et al. Pioglitazone-incorporated nanoparticles prevent plaque destabilization and rupture by regulating monocyte/macrophage differentiation in ApoE-/- mice. *Arterioscler Thromb Vasc Biol.* 2016; 36:491–500. doi: 10.1161/ATVBAHA.115.307057.
- Nemati S, Kim SJ, Shin YM, Shin H. Current progress in application of polymeric nanofibers to tissue engineering. *Nano Converg.* 2019; 6: 36. doi: 10.1186/s40580-019-0209-y.
- Noukeu LC, Wolf J, Yuan B, Banerjee S, Nguyen KT. Nanoparticles for detection and treatment of peripheral arterial disease. *Small.* 2018; 14:e1800644. doi: 10.1002/sml.201800644. Crossref Medline Google Scholar
- Ovcharenko EA, Seifalian A, Rezvova MA et al. A New Nanocomposite Copolymer Based On Functionalised Graphene Oxide for Development of Heart Valves. *Sci Rep.* 2020; 10: 5271. doi.org/10.1038/s41598-020-62122-8.
- Paszkwicz S, Pawlikowska D, Szymczyk A et al. Interfacial interactions in PTT–PTMO/polyhedral oligomeric silsesquioxane (POSS) nanocomposites and their impact on mechanical, thermal, and dielectric properties. *Polym Bull.* 2018; 75:4999–5014. https://doi.org/10.1007/s00289-018-2317-y.
- Pérez-Medina C, Binderup T, Lobatto ME, et al. In vivo PET imaging of HDL in multiple atherosclerosis models. *JACC Cardiovasc Imaging.* 2016; 9:950–961. doi: 10.1016/j.jcmg.2016.01.020.
- Reed ML, Wu C, Kneller J et al. Micromechanical devices for intravascular drug delivery. *J Pharm Sci.* 1998; 87: 1387-1394. doi.org/10.1021/js980085q.
- Ridker PM, Everett BM, Thuren T et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease: lessons from the CANTOS trial. *N Engl J Med.* 2017; 377:1119–1131. doi: 10.1056/NEJMoa1707914.
- Rippel RA, Ghanbari H, Seifalian AM. Tissue-engineered heart valve: future of cardiac surgery. *World J. Surg.* 36, 2012; 36(7):1581-1591. doi: 10.1007/s00268-012-1535-y.
- Rose, J.C.; De Laporte, L. Hierarchical Design of Tissue Regenerative Constructs. *Adv. Healthc Mater.* 2018; 7: e1701067. doi: 10.1002/adhm.201701067.
- Rudd JH, Myers KS, Bansilal S et al. Atherosclerosis inflammation imaging with 18F-FDG PET: carotid, iliac, and femoral uptake reproducibility, quantification methods, and recommendations. *J Nucl Med.* 2008; 49:871–878. doi: 10.2967/jnumed.107.050294.
- Sager HB, Dutta P, Dahlman JE, et al. RNAi targeting multiple cell adhesion molecules reduces immune cell recruitment and vascular inflammation after myocardial infarction. *Sci Transl Med.* 2016; 8:342ra80. doi: 10.1126/scitranslmed.aaf1435.
- Seedial SM, Ghosh S, Saunders RS et al. Local drug delivery to prevent restenosis. *J Vasc Surg.* 2013; 57:1403–1414. doi: 10.1016/j.jvs.2012.12.069.
- Seijkens TTP, van Tiel CM, Kusters PJH et al. Targeting CD40-induced TRAF6 signaling in macrophages reduces atherosclerosis. *J Am Coll Cardiol.* 2018;71:527–542. doi: 10.1016/j.jacc.2017.11.055.
- Seijkens TTP, van Tiel CM, Kusters PJH et al. Targeting CD40-induced TRAF6 signaling in macrophages reduces atherosclerosis. *J Am Coll Cardiol.* 2018; 71:527–542. doi: 10.1016/j.jacc.2017.11.055.
- Shi J, Kantoff PW, Wooster R, Farokhzad OC. Cancer nanomedicine: progress, challenges and opportunities. *Nat Rev Cancer.* 2017;17:20–37. doi: 10.1038/nrc.2016.108.
- Smith BR, Gambhir SS. Nanomaterials for in vivo imaging. *Chem Rev.* 2017;117:901–986. doi: 10.1021/acs.chemrev.6b00073.
- Smith BR, Ghosn EE, Rallapalli H et al. Selective uptake of single-walled carbon nanotubes by circulating monocytes for enhanced tumour delivery. *Nat Nanotechnol.* 2014; 9:481–487. doi: 10.1038/nnano.2014.62.
- Tadin-Strapps M, Peterson LB, Cumiskey AM et al. siRNA-induced liver ApoB knockdown lowers serum LDL-cholesterol in a mouse model with human-like serum lipids. *J Lipid Res.* 2011; 52:1084–1097. doi: 10.1194/jlr.M012872.
- Takahata K, Gianchandani KT. A planar approach for manufacturing cardiac stents: design, fabrication, and mechanical evaluation. *J Microelectromech Syst.* 2004;13: 933-939. doi: 10.1109/JMEMS.2004.838357.

- Tang J, Lobatto ME, Hassing L et al. Inhibiting macrophage proliferation suppresses atherosclerotic plaque inflammation. *Sci Adv*. 2015; 1:e1400223. doi: 10.1126/sciadv.1400223.
- Tsukie N, Nakano K, Matoba T et al. Pitavastatin-incorporated nanoparticle-eluting stents attenuate in-stent stenosis without delayed endothelial healing effects in a porcine coronary artery model. *J Atheroscler Thromb*. 2013; 20:32–45. doi: 10.5551/jat.13862.
- Tsukie N, Nakano K, Matoba T et al. Pitavastatin-incorporated nanoparticle-eluting stents attenuate in-stent stenosis without delayed endothelial healing effects in a porcine coronary artery model. *J Atheroscler Thromb*. 2013; 20:32–45. doi: 10.5551/jat.13862.
- Uwatoku T, Shimokawa H, Abe K et al. Application of nanoparticle technology for the prevention of restenosis after balloon injury in rats. *Circ Res*. 2003; 92: 62-69. doi:10.1161/01.res.0000069021.56380.e2.
- Van der Valk FM, van Wijk DF, Lobatto ME et al. Prednisolone-containing liposomes accumulate in human atherosclerotic macrophages upon intravenous administration. *Nanomedicine*. 2015; 11:1039–1046. doi: 10.1016/j.nano.2015.02.021.
- Ventola CL. Progress in nanomedicine: approved and investigational nanodrugs. *P T*. 2017; 42:742–755.
- Virmani R, Guagliumi G, Farb A et al. Localized hypersensitivity and late coronary thrombosis secondary to a sirolimus-eluting stent: should we be cautious? *Circulation*. 2004; 109: 701-705. doi: 10.1161/01.CIR.0000116202.41966.D4.
- Wang D, Xu Y, Li Q, Turng LS. Artificial small-diameter blood vessels: Materials, fabrication, surface modification, mechanical properties, and bioactive functionalities. *J Mater. Chem*. 2020; 8: 1801–1822. doi: 10.1039/c9tb01849b.
- Wang Y, Li L, Zhao W et al. Targeted therapy of atherosclerosis by a broad-spectrum reactive oxygen species scavenging nanoparticle with intrinsic anti-inflammatory activity. *ACS Nano*. 2018; 12:8943–8960. doi: 10.1021/acsnano.8b02037.
- Wang Z, Wu Y, Wang J et al. Effect of Resveratrol on Modulation of Endothelial Cells and Macrophages for Rapid Vascular Regeneration from Electrospun Poly(ϵ -caprolactone) Scaffolds *ACS Appl Mater Interfaces*. 2017; 9: 19541-19551. doi: 10.1021/acsmi.6b16573.
- Winter PM, Neubauer AM, Caruthers SD et al. Endothelial alpha(v)beta3 integrin-targeted fumagillin nanoparticles inhibit angiogenesis in atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2006; 26:2103–2109. doi: 10.1161/01.ATV.0000235724.11299.76
- World Health Organization. The Top 10 Causes of Death.2018. <https://www.who.int/news-room/fact-sheets/detail/the-top-10-causes-of-death> Accessed November 4, 2018
- Yeo GC. A New Vascular Engineering Strategy Using 3D Printed Ice. *Trends Biotechnol*. 2019; 37: 451–453. doi: 10.1016/j.tibtech.2019.01.007.
- Yin RX, Yang DZ, Wu JZ. Nanoparticle drug- and gene-eluting stents for the prevention and treatment of coronary restenosis. *Theranostics*. 2014; 4:175–200. doi: 10.7150/thno.7210.
- Yu M, Amengual J, Menon A et al. Targeted nanotherapeutics encapsulating liver X receptor agonist GW3965 enhance antiatherogenic effects without adverse effects on hepatic lipid metabolism in Ldlr(-/-) mice. *Adv Healthc Mater*. 2017; 6. doi: 10.1002/adhm.201700313. doi: 10.1002/adhm.201700313.
- Zagho MM, Hussein EA, Elzatahry AA. Recent Overviews in Functional Polymer Composites for Biomedical Applications. *Polymers (Basel)*. 2018; 10: 739 doi: 10.3390/polym10070739.
- Zimmermann TS, Lee AC, Akinc A et al. RNAi-mediated gene silencing in non-human primates. *Nature*. 2006; 441:111–114. doi: 10.1038/nature04688.
